

FIG. 1A

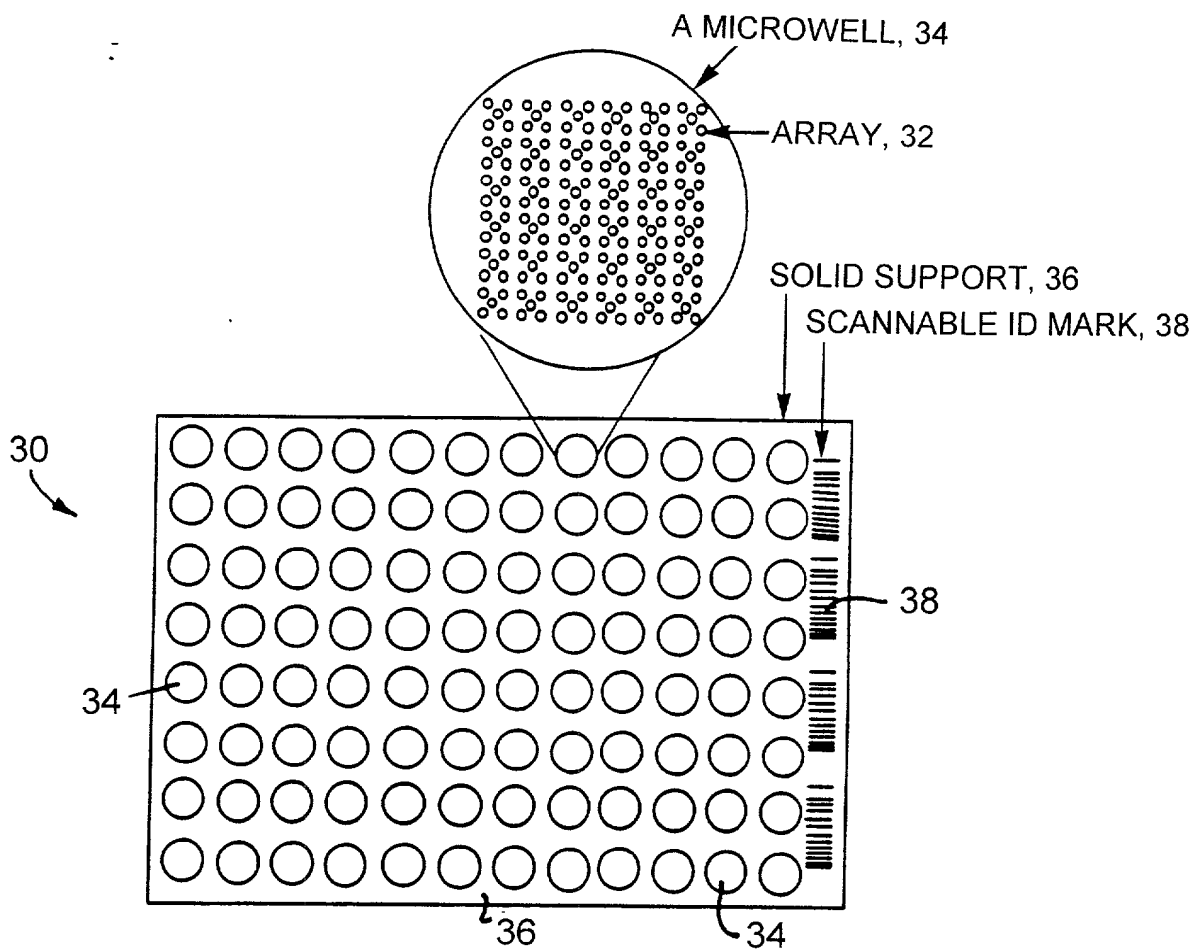


FIG. 1B

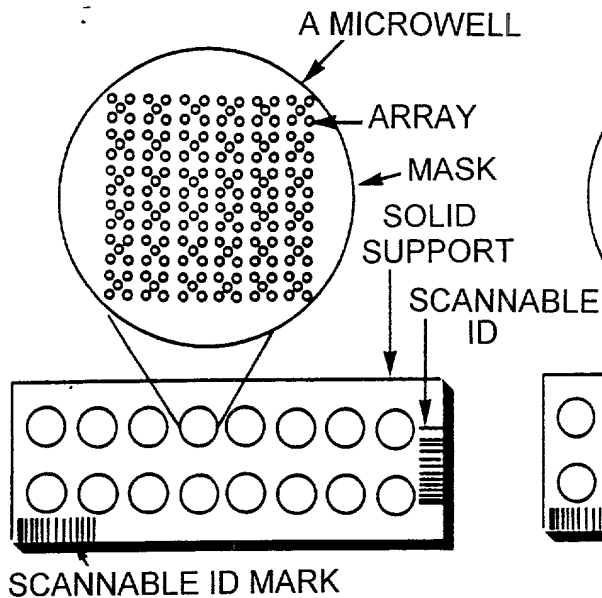


FIG. 2A

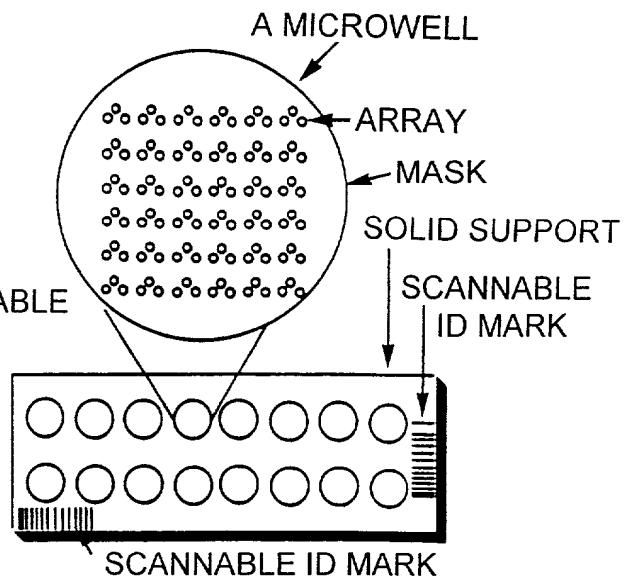


FIG. 2B

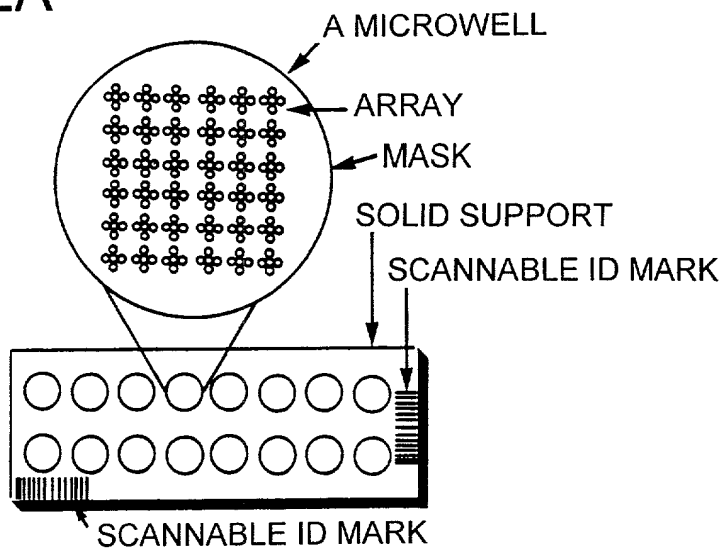
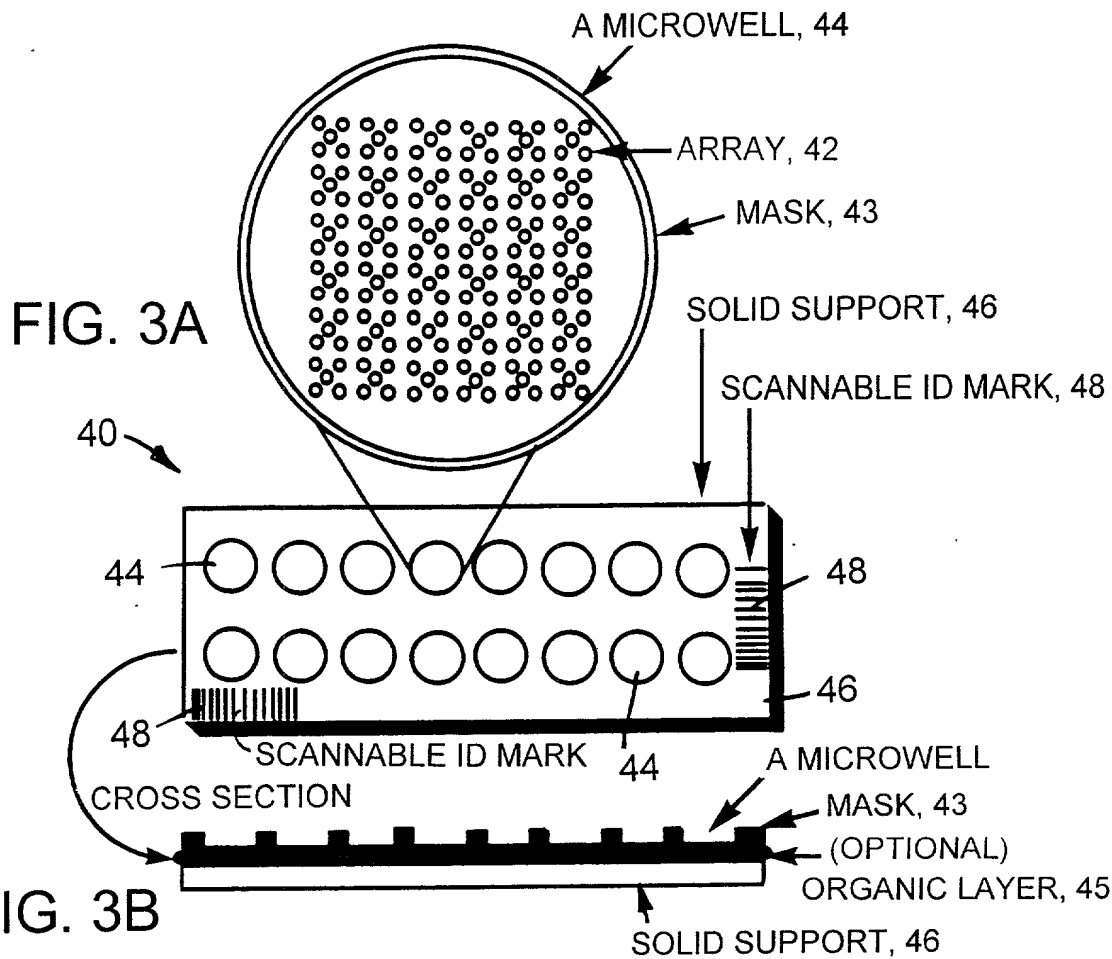
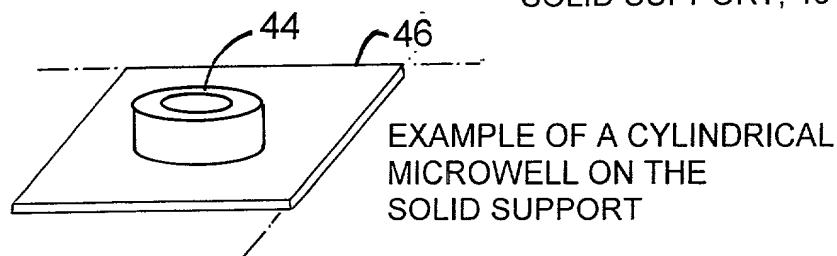


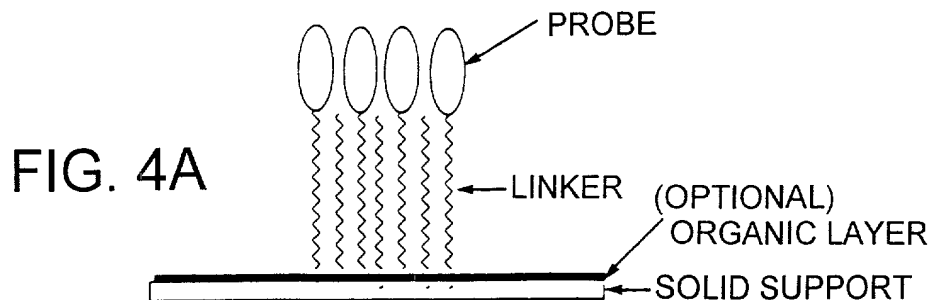
FIG. 2C

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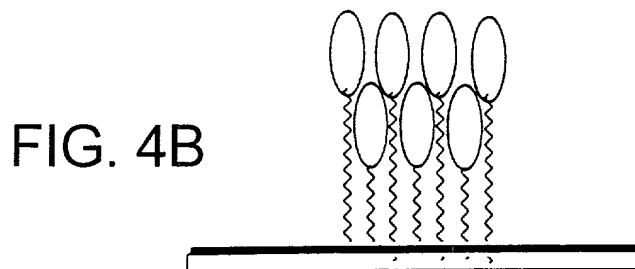


**FIG. 3B**

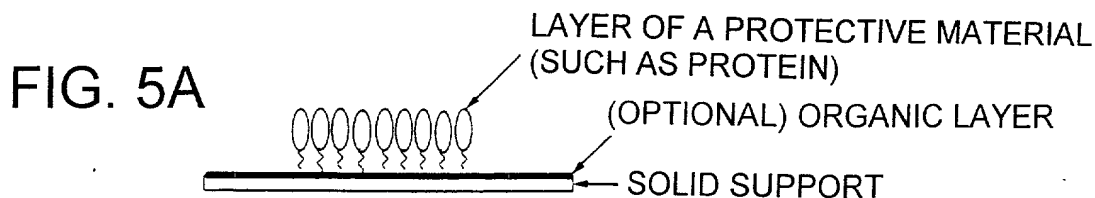




ATTACHING PROBES TO SOLID-SUPPORT WITH LINKERS OF THE SAME LENGTH RESULTS IN LOWER DENSITY OF PROBE ATTACHMENT



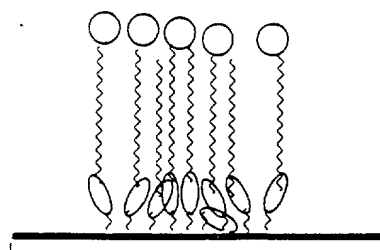
ATTACHING PROBES TO SOLID-SUPPORT WITH LINKERS OF DIFFERENT LENGTHS RESULTS IN MUCH HIGHER DENSITY OF PROBE ATTACHMENT



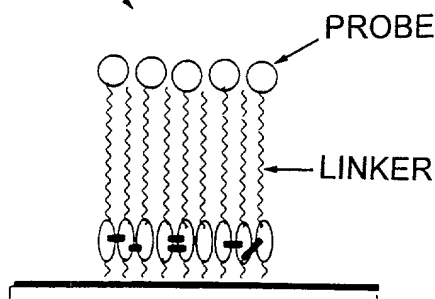
A LAYER OF PROTECTIVE MATERIAL IS LAID WITH SMALL LINKERS

NO CROSS-LINKING

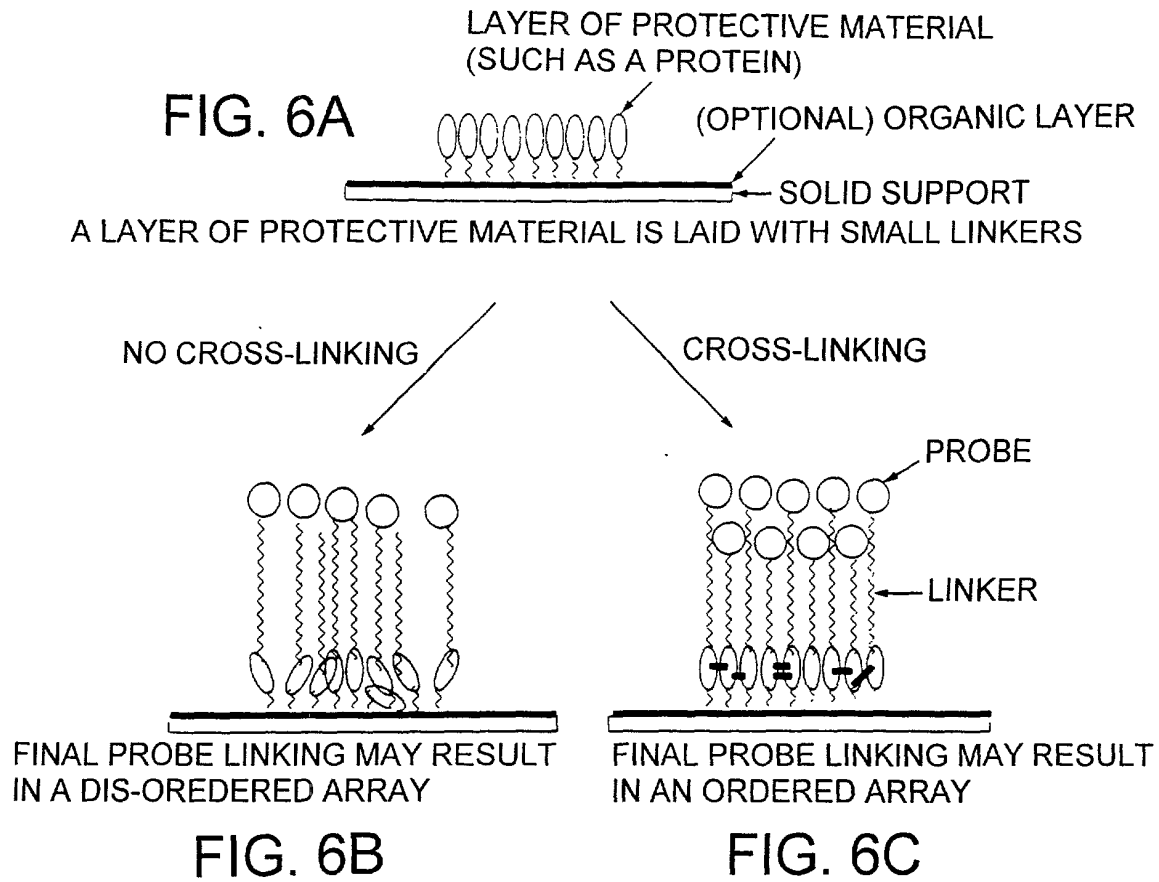
CROSS-LINKING

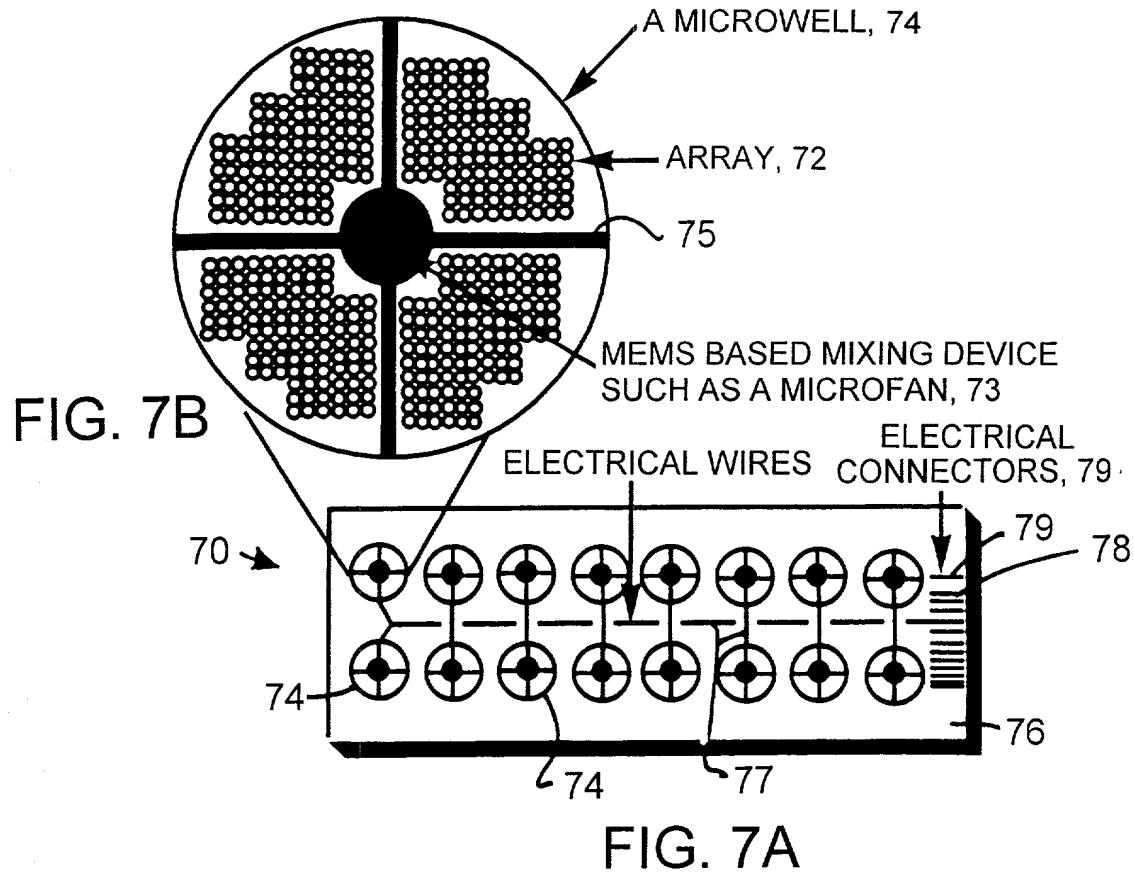


FINAL PROBE LINKING MAY RESULT IN A DIS-ORDERING ARRAY



FINAL PROBE LINKING MAY RESULT IN AN ORDERED ARRAY





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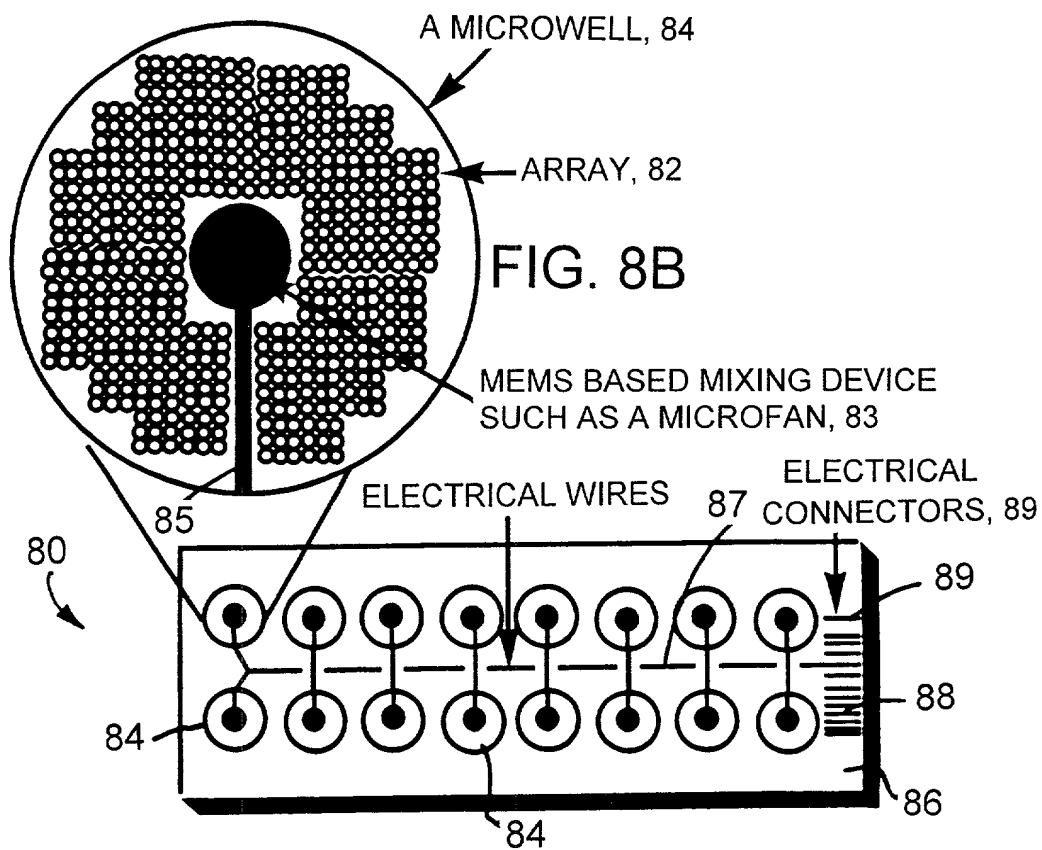
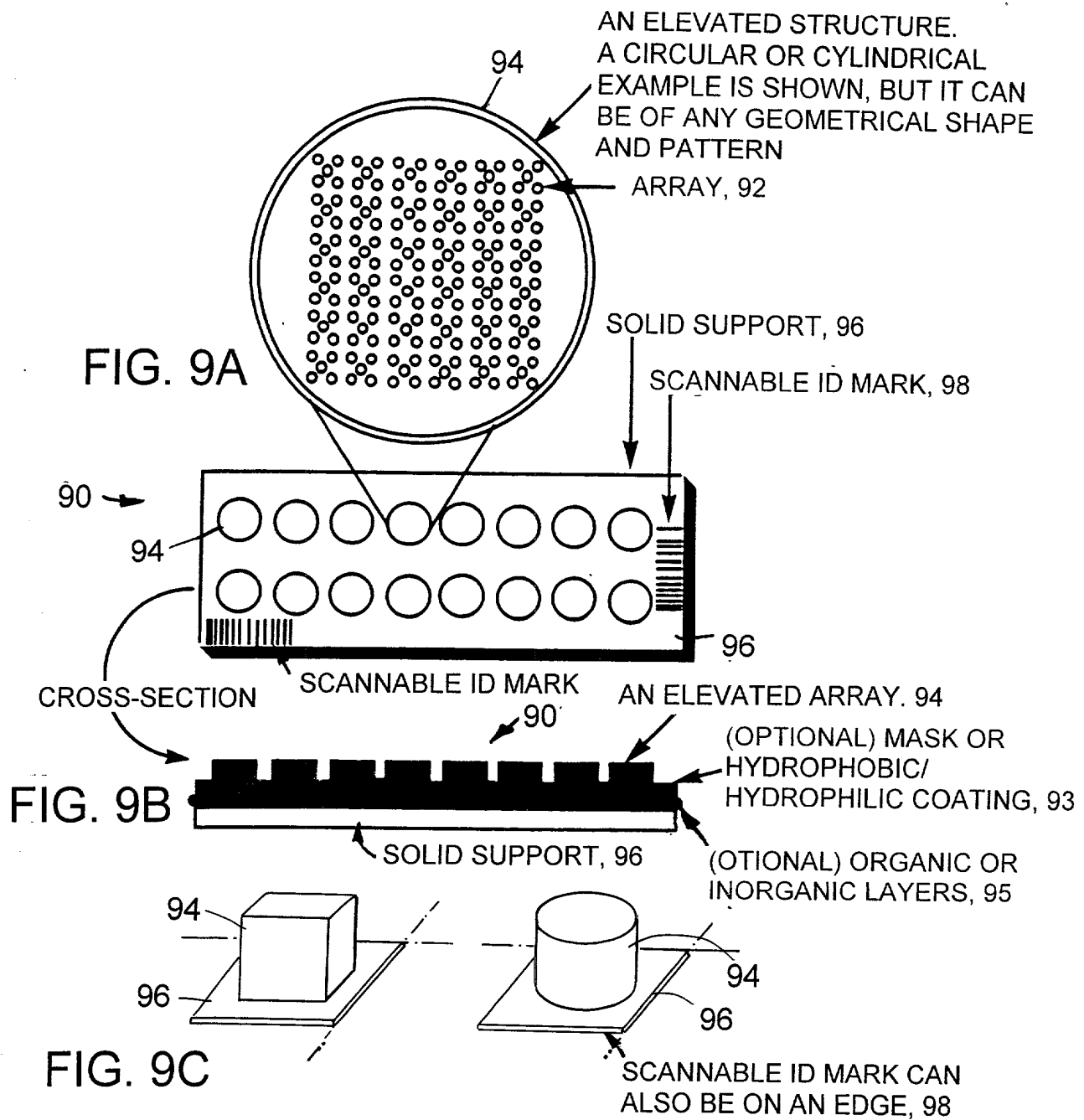


FIG. 8A





EXAMPLES OF ELEVATED STRUCTURES ON SOLID SUPPORT.  
 THE STRUCTURES CAN BE CYLINDRICAL OR CUBOID OR ANY  
 OTHER GEOMETRICAL SHAPE

EACH ELEVATED STRUCTURE  
CONTAINS A PROBE ARRAY

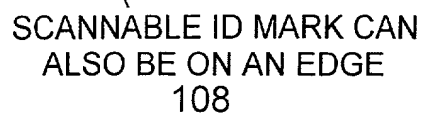


FIG. 1 is a perspective view of a substrate 100. The substrate 100 is a rectangular plate with a thickness 106. On the top surface of the substrate 100, there are four circular features 104 arranged in a 2x2 grid. An arrow points to the top surface of the substrate 100.

100 →

106

104

SCANNABLE ID MARK CAN ALSO BE ON AN EDGE

108

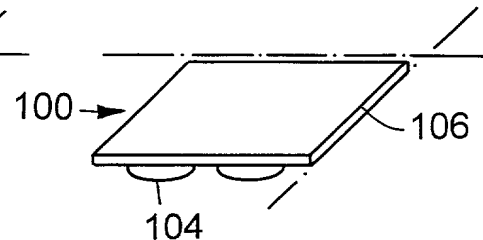
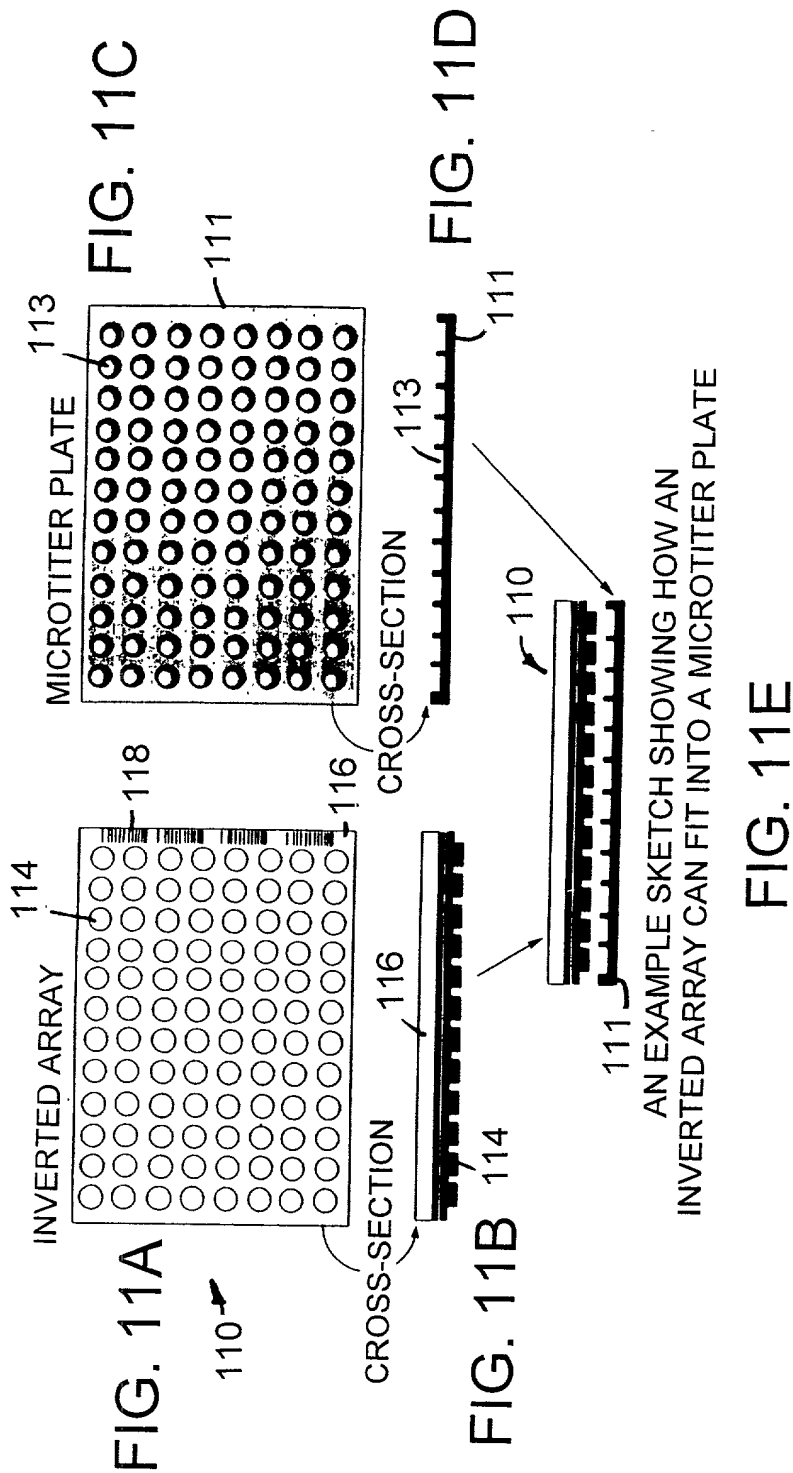
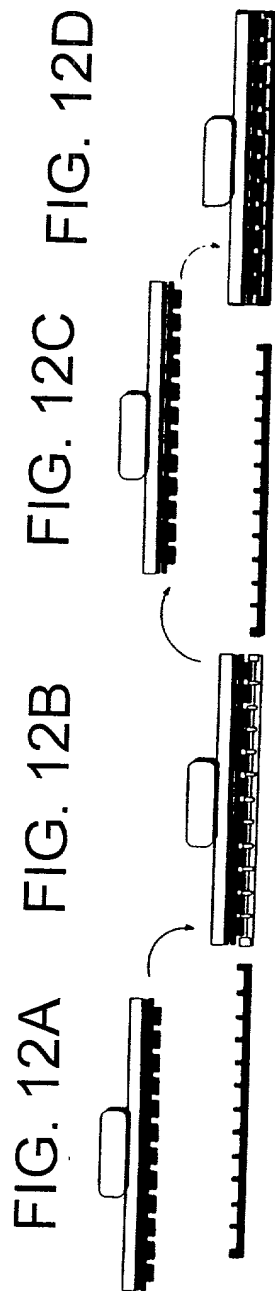


Diagram illustrating a system for result read-out. The system includes a base (100) supporting a structure (104) containing capillaries or optical fibers or electrical contacts (105). The structure is connected to a system for result read-out (106) via a connection (107).

FIG. 10C





THE "INVERTED ARRAY" CAN BE MOVED FROM ONE REACTION VESSEL TO ANOTHER. IT CAN BE MOVED EITHER USING A BUILT-IN HANDLE OR USING A VACUUM SUCTION DEVICE OR ANY OTHER MECHANISM. THE PROCESS CAN BE DONE MANUALLY OR ROBOTICALLY AND THE ASSAY PROCEDURE CAN BE EASILY AUTOMATED. THIS SCHEMATIC SHOWS AN EXAMPLE OF ONE SUCH PROCESS.

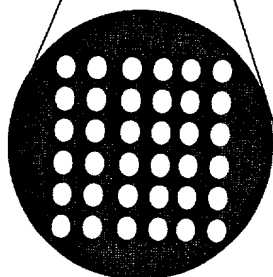
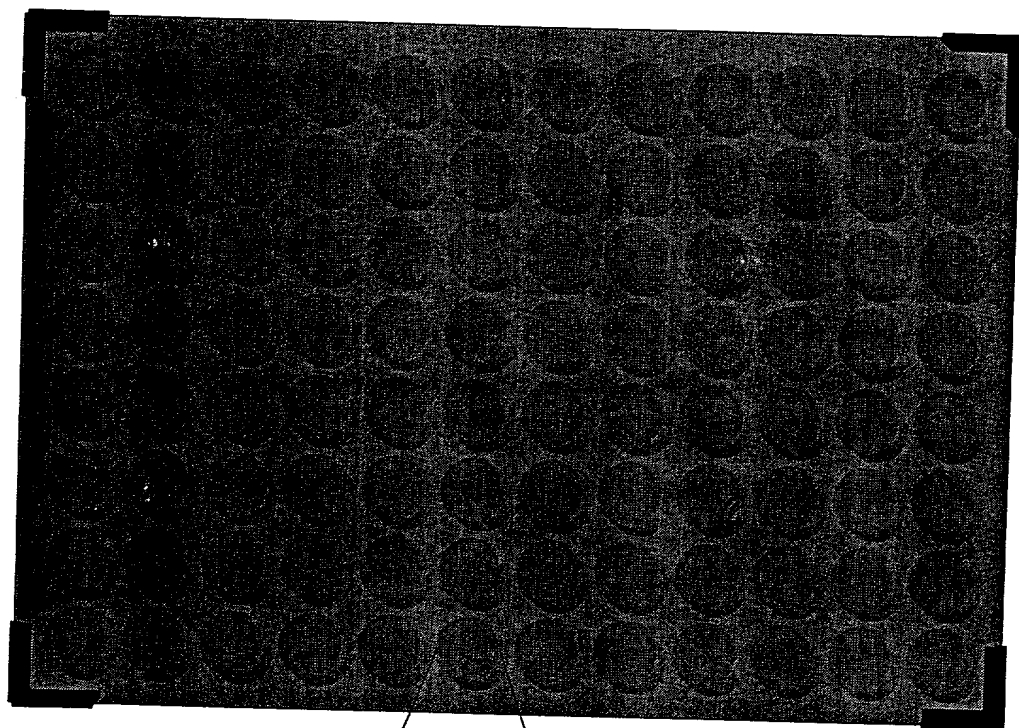


FIG. 13A

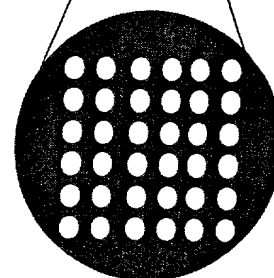
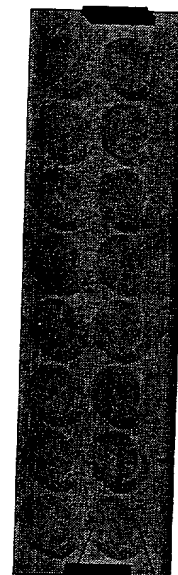


FIG. 13B

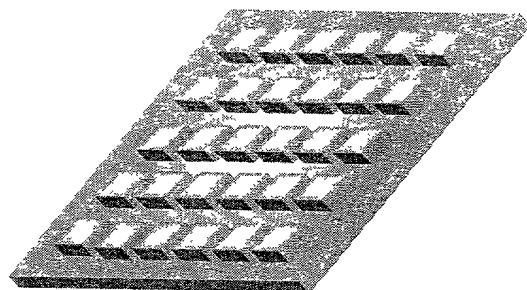


FIG. 13C

208220" 99096660

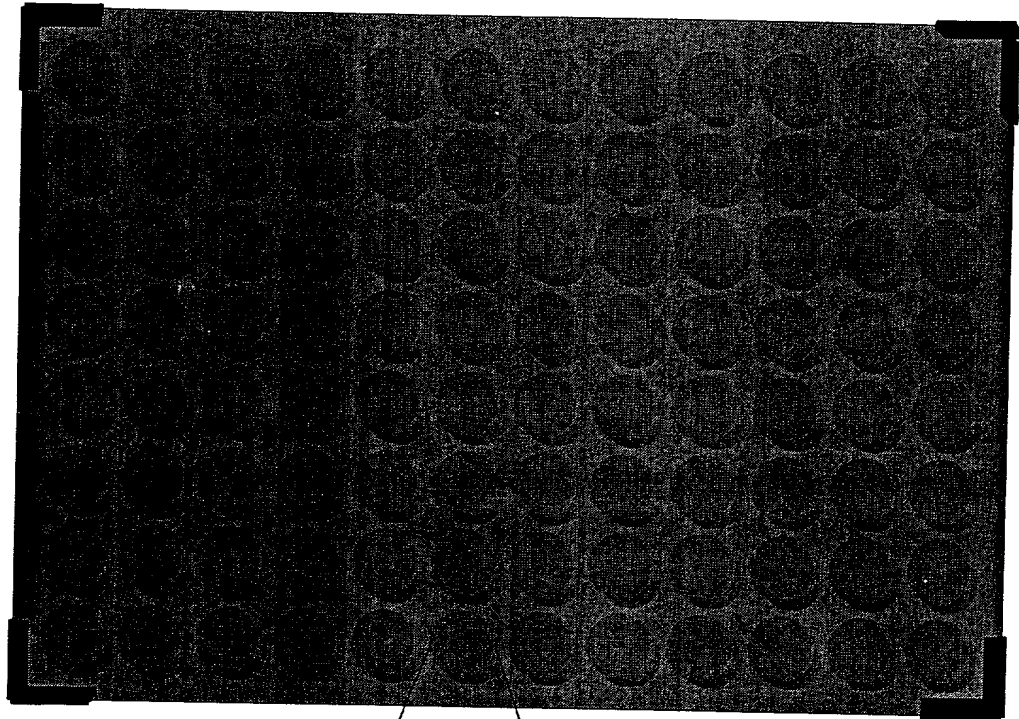


FIG. 14A

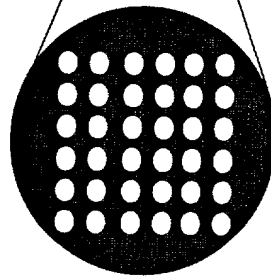


FIG. 14B Elevated sub-structure



FIG. 14C Planar sub-structure

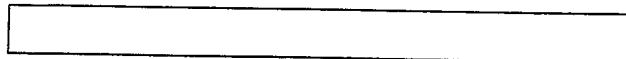
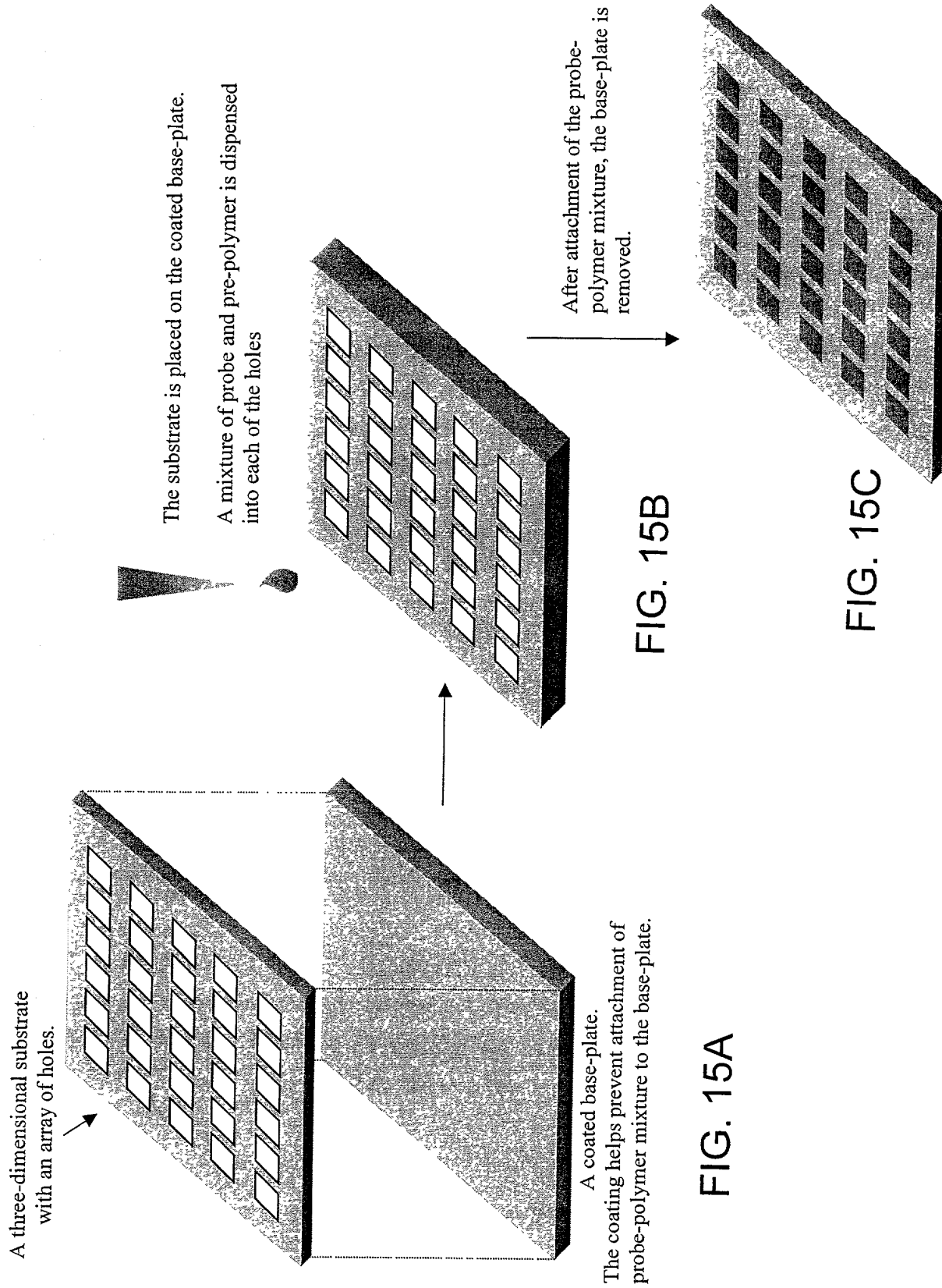


FIG. 14D Depressed sub-structure





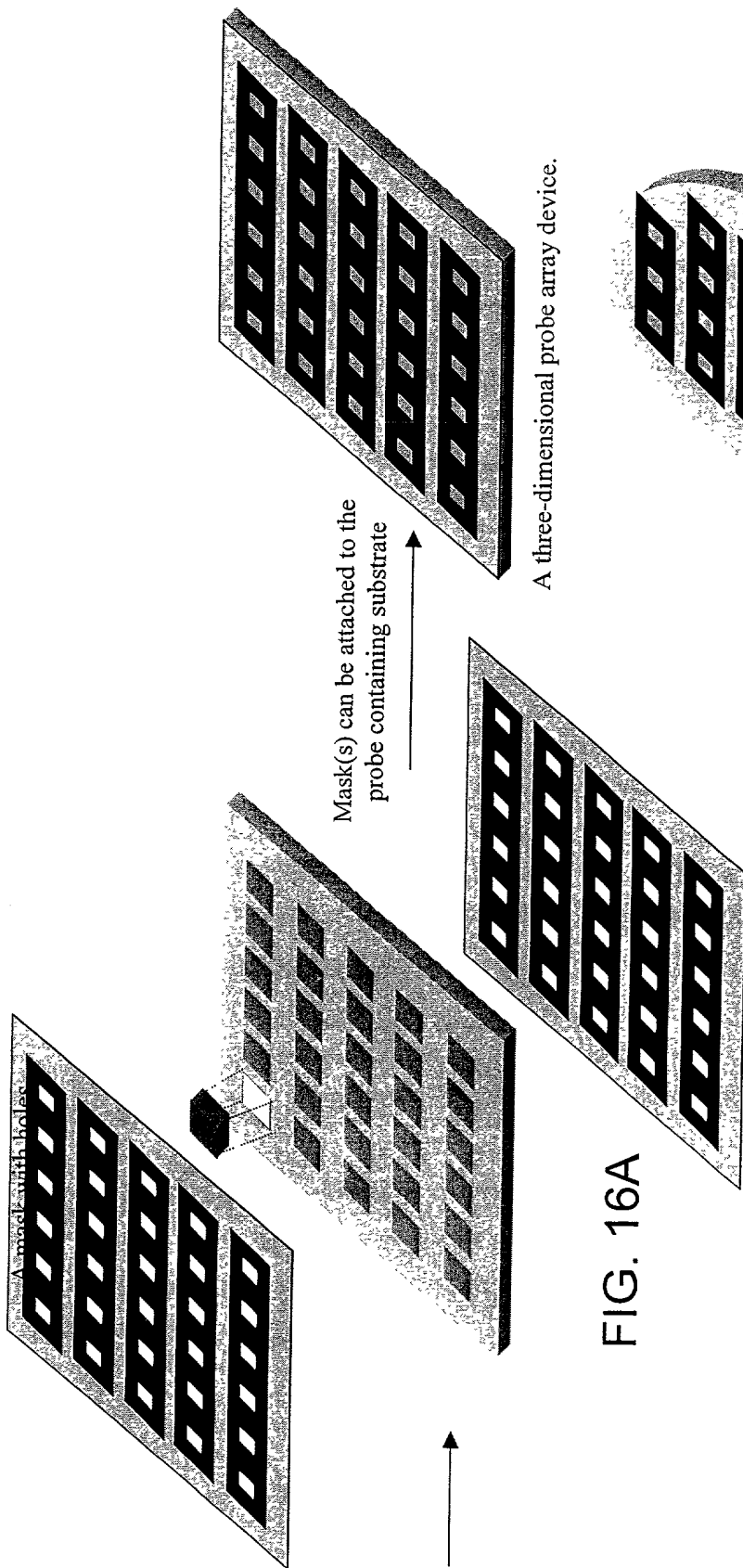


FIG. 16A

A three-dimensional probe array device.

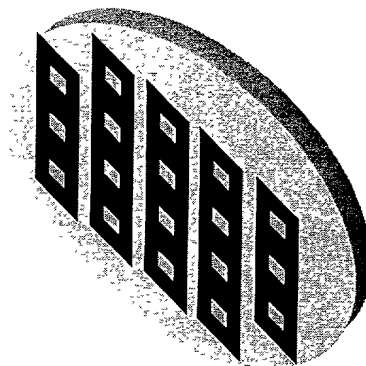


FIG. 16B

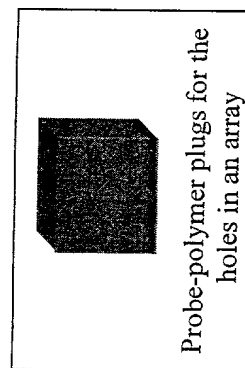


FIG. 16C



The microarray biochip can  
Also be housed in a sealed chamber.

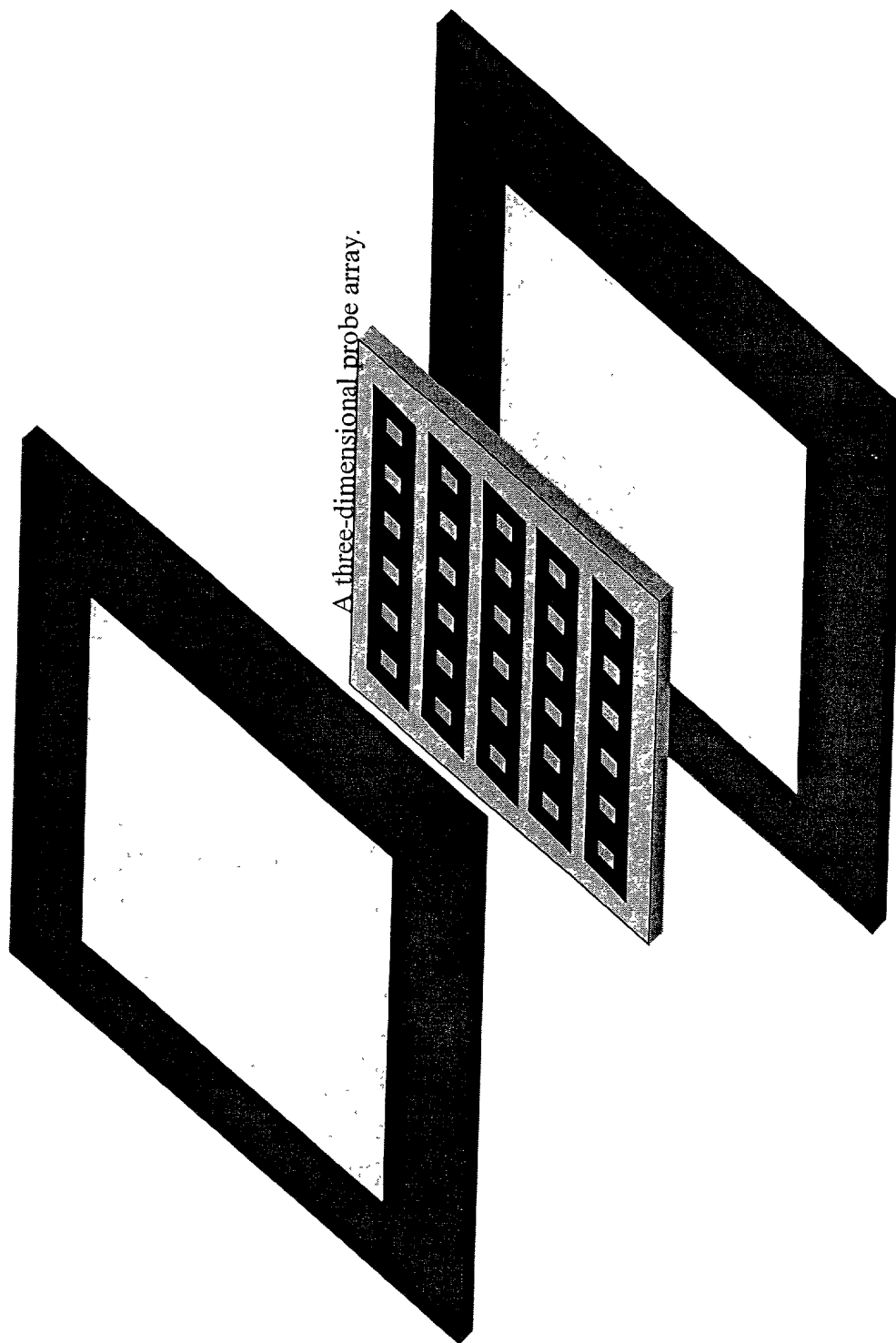
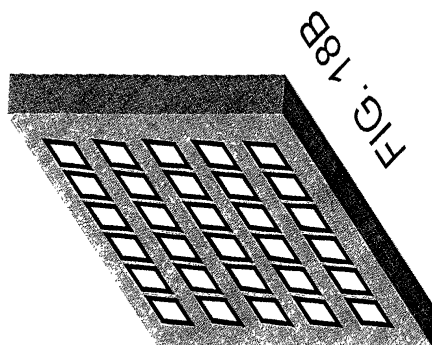
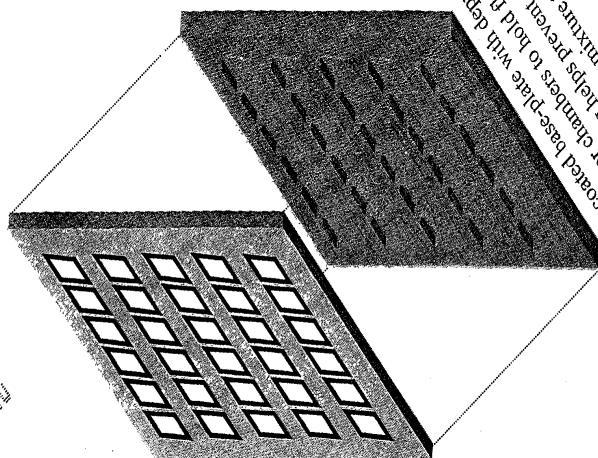


FIG. 17



20220-959640



The coating helps prevent attachment of probe-polymer mixture to the base-plate.

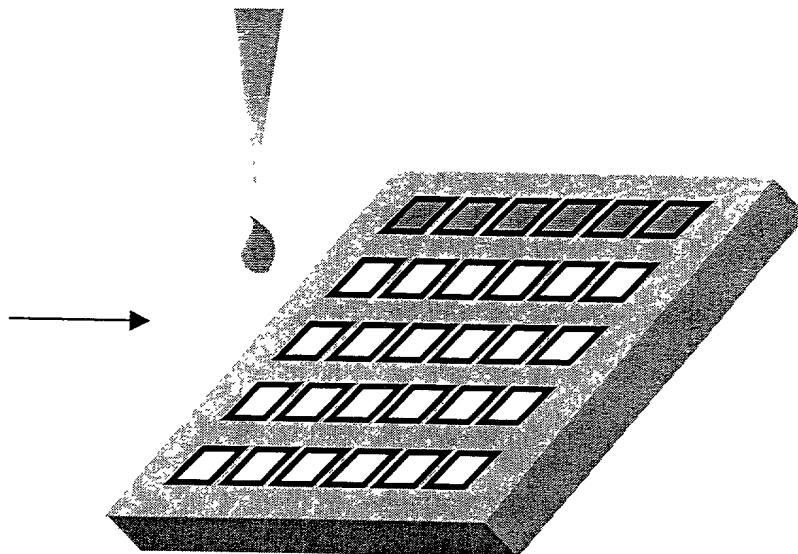
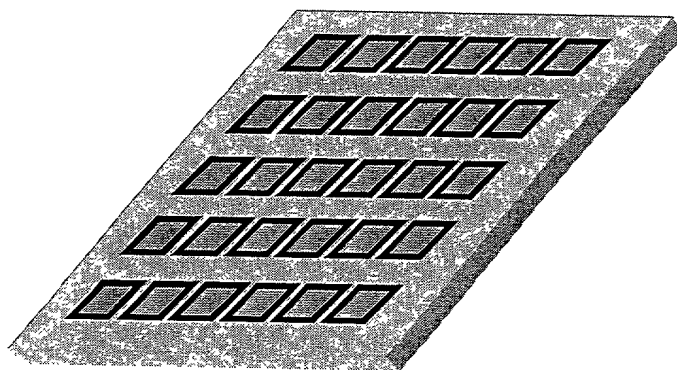


FIG. 19A



Another implementation of 3D porous biochip

FIG. 19B

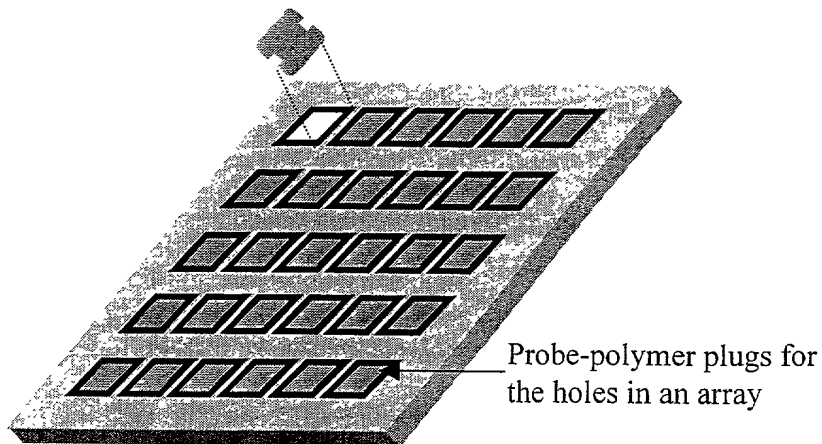


FIG. 19C

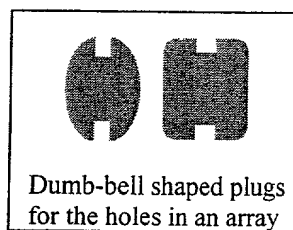
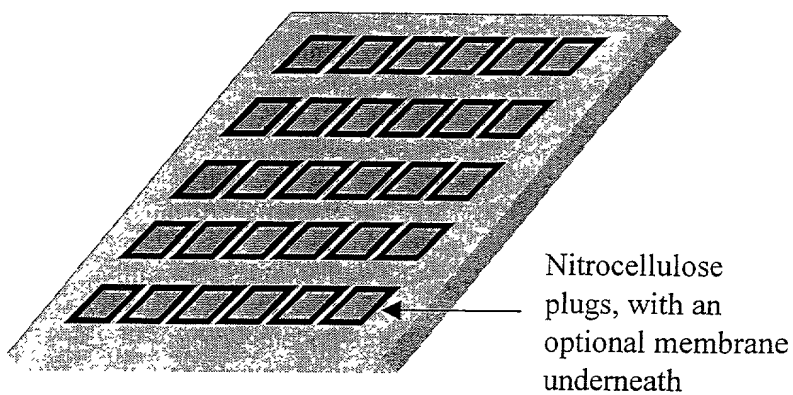


FIG. 19D



Two examples of the types of material that can be used to manufacture the 3D porous array

FIG. 19E

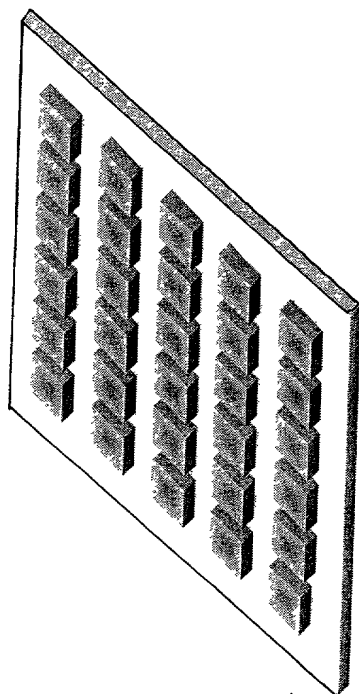


FIG. 20A

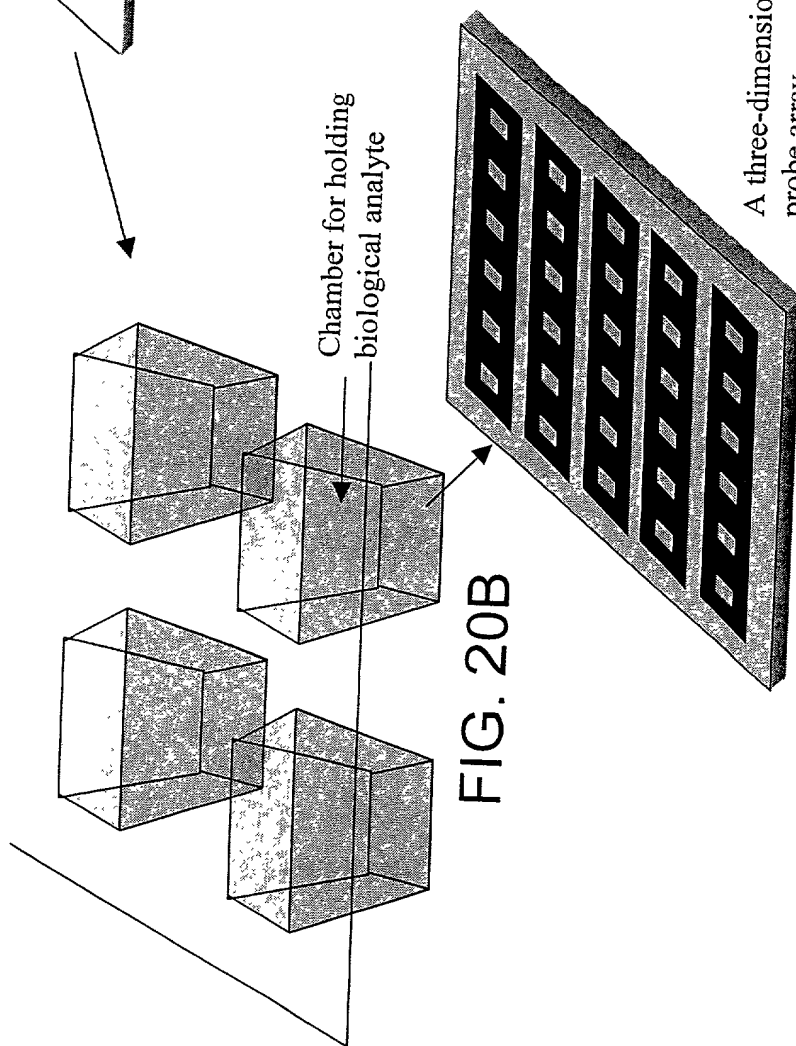
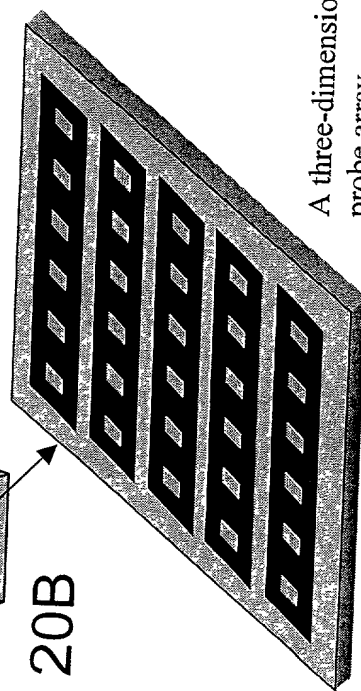


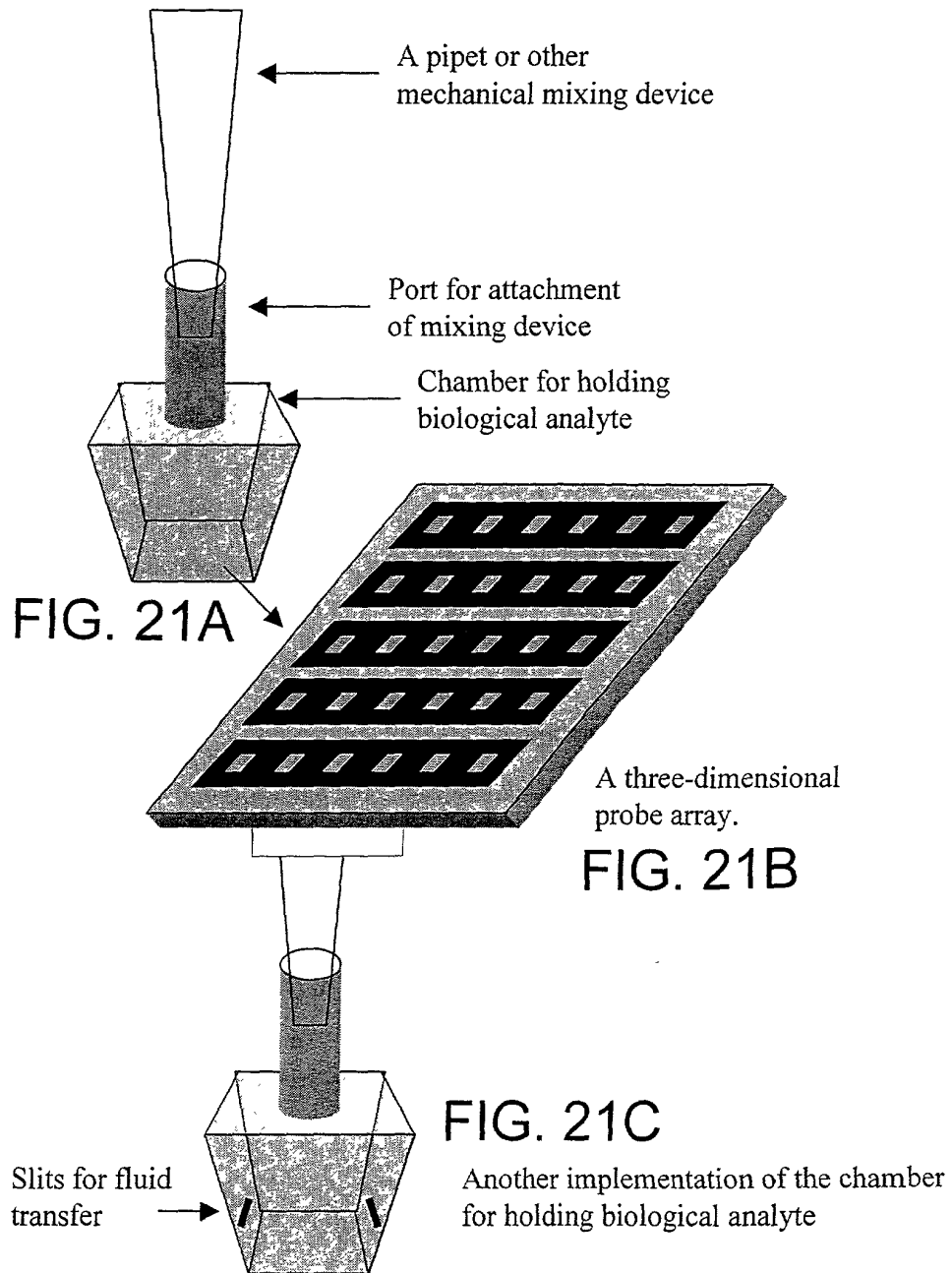
FIG. 20B

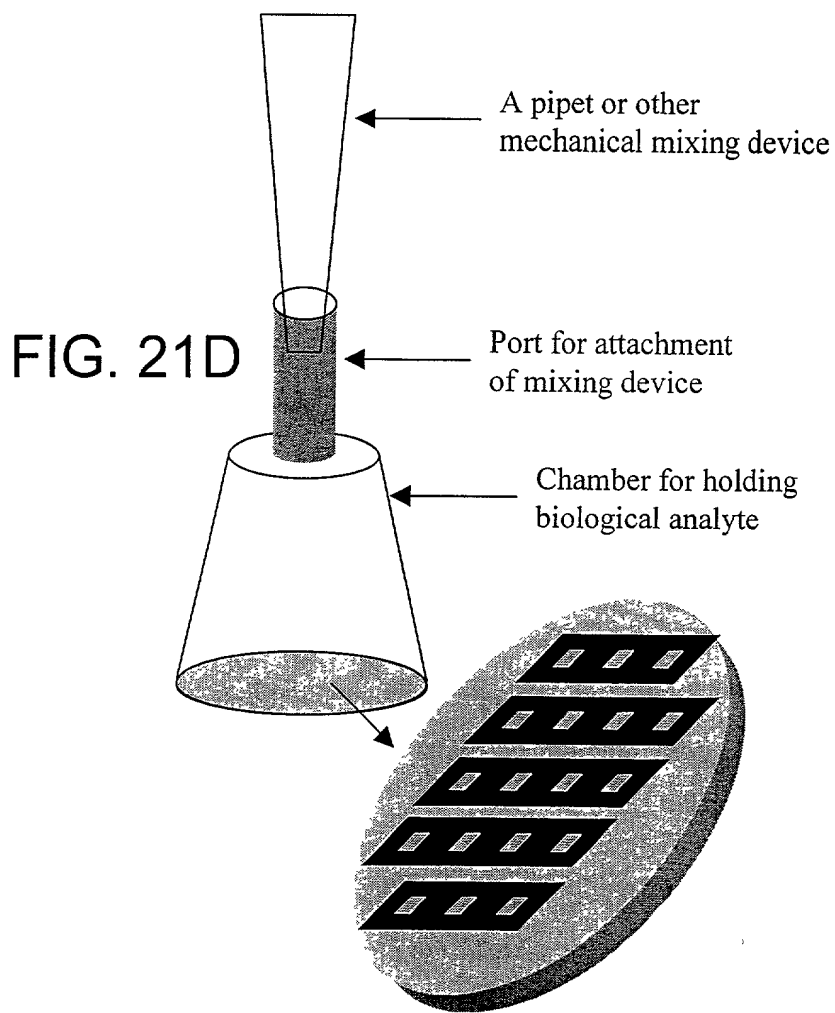


A three-dimensional probe array.

FIG. 20C

The microarray biochip can also be housed in a sealed hand-held or Point of Care device.





A three-dimensional probe array.

FIG. 21E

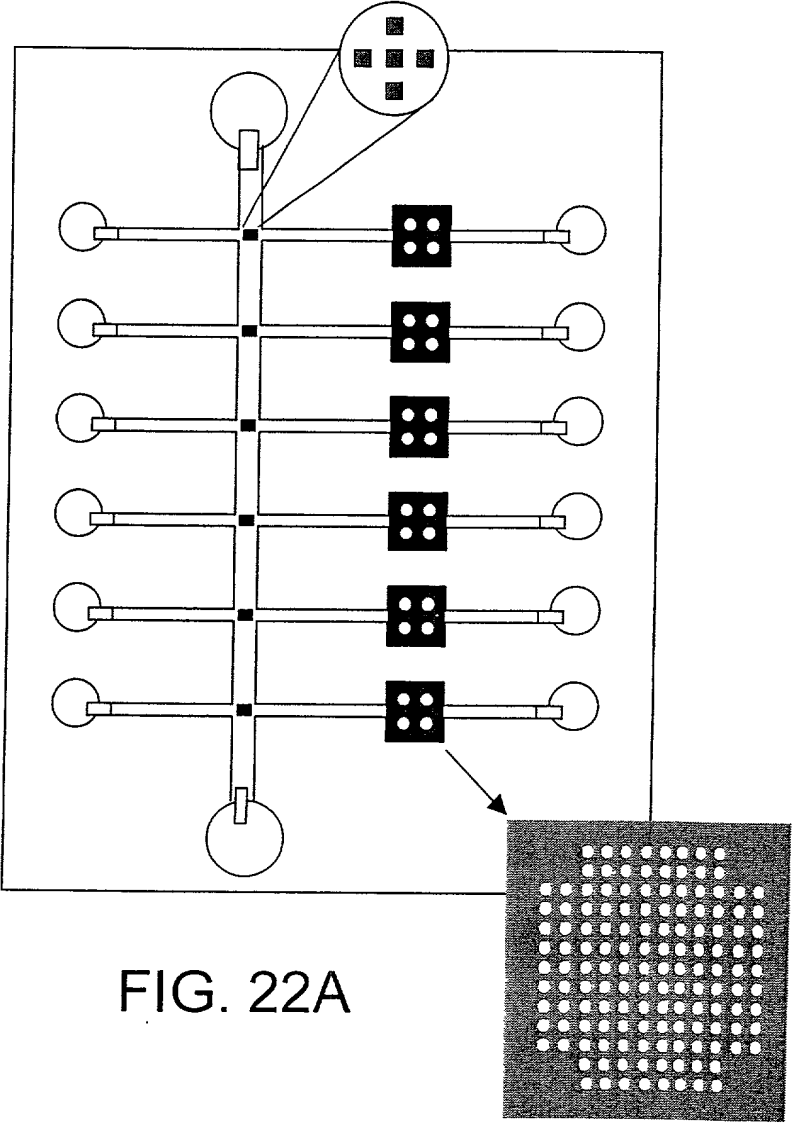


FIG. 22A



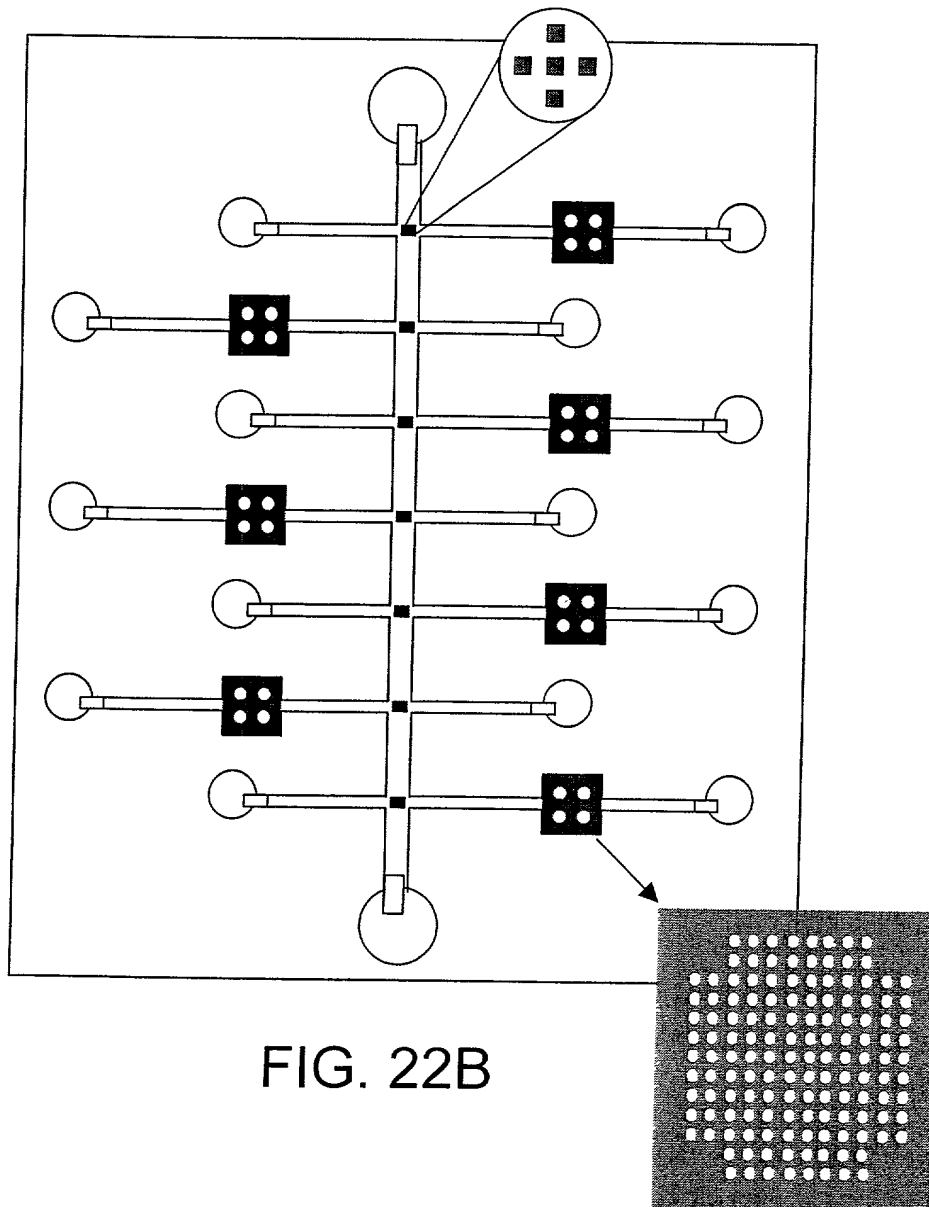


FIG. 22B

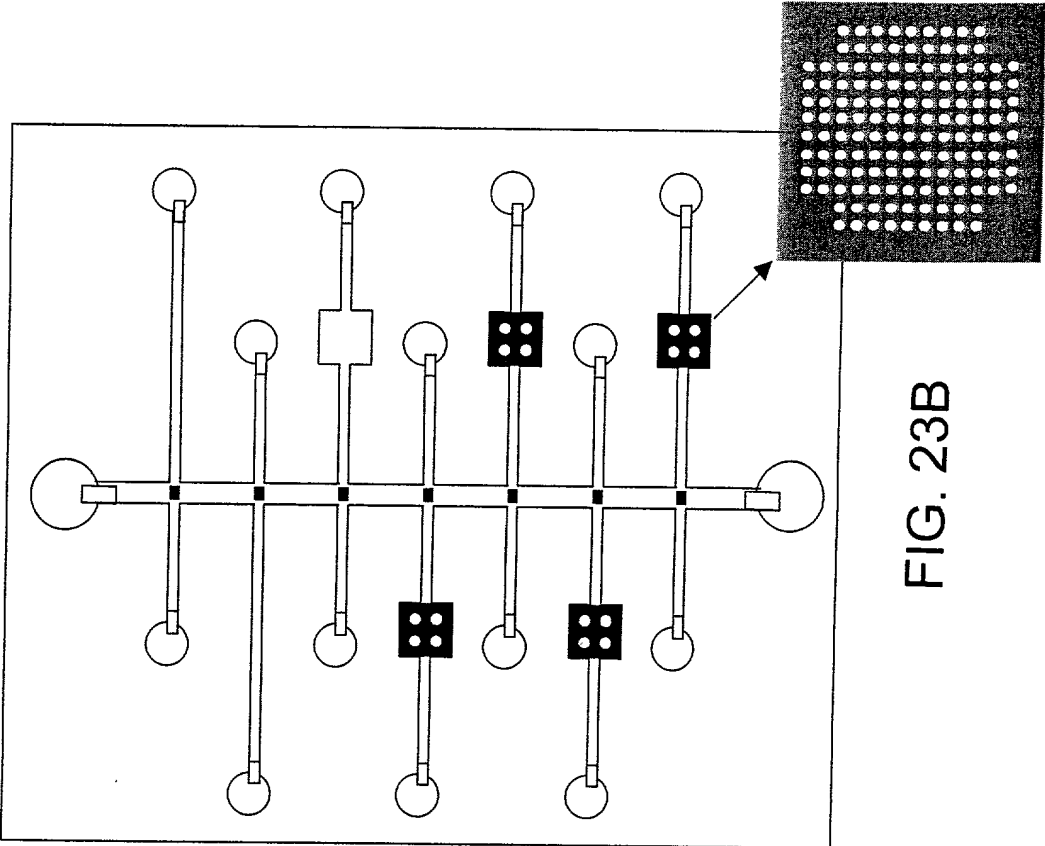


FIG. 23B

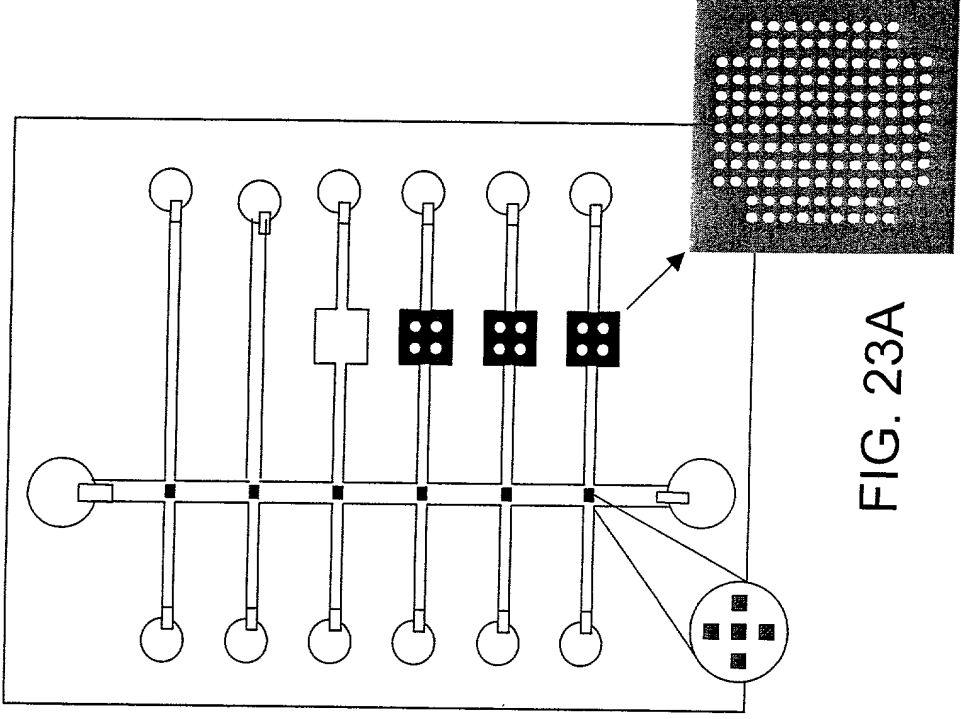
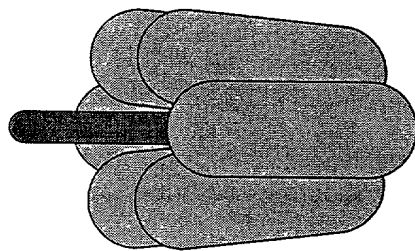


FIG. 23A

# ATPase based Fluid-Micromixers



Cartoon of the multi-subunit enzyme - ATPase - that rotates in response to ATP synthesis to hydrolysis. The centrally located  $\gamma$ -subunit rotates relative to the hexameric  $\alpha, \beta$ -subunit core.

FIG. 24A

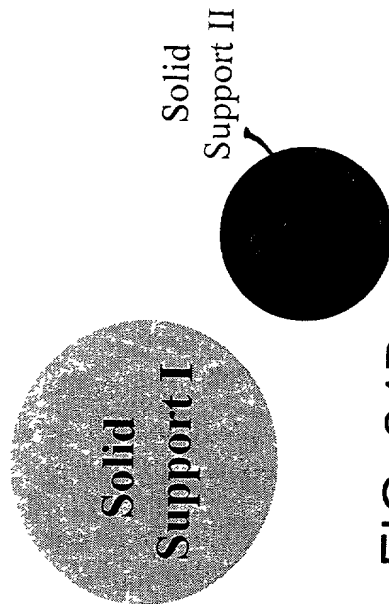
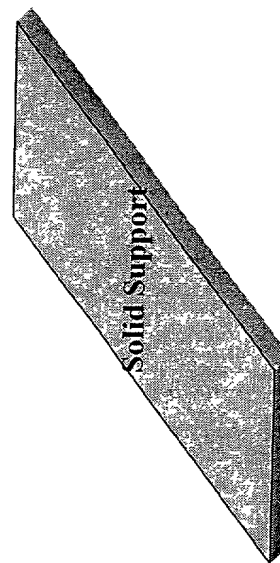


FIG. 24B

Spherical Beads as Solid supports for attaching different subunits of ATPase

Linkers for attaching different subunits of ATPase to the solid support

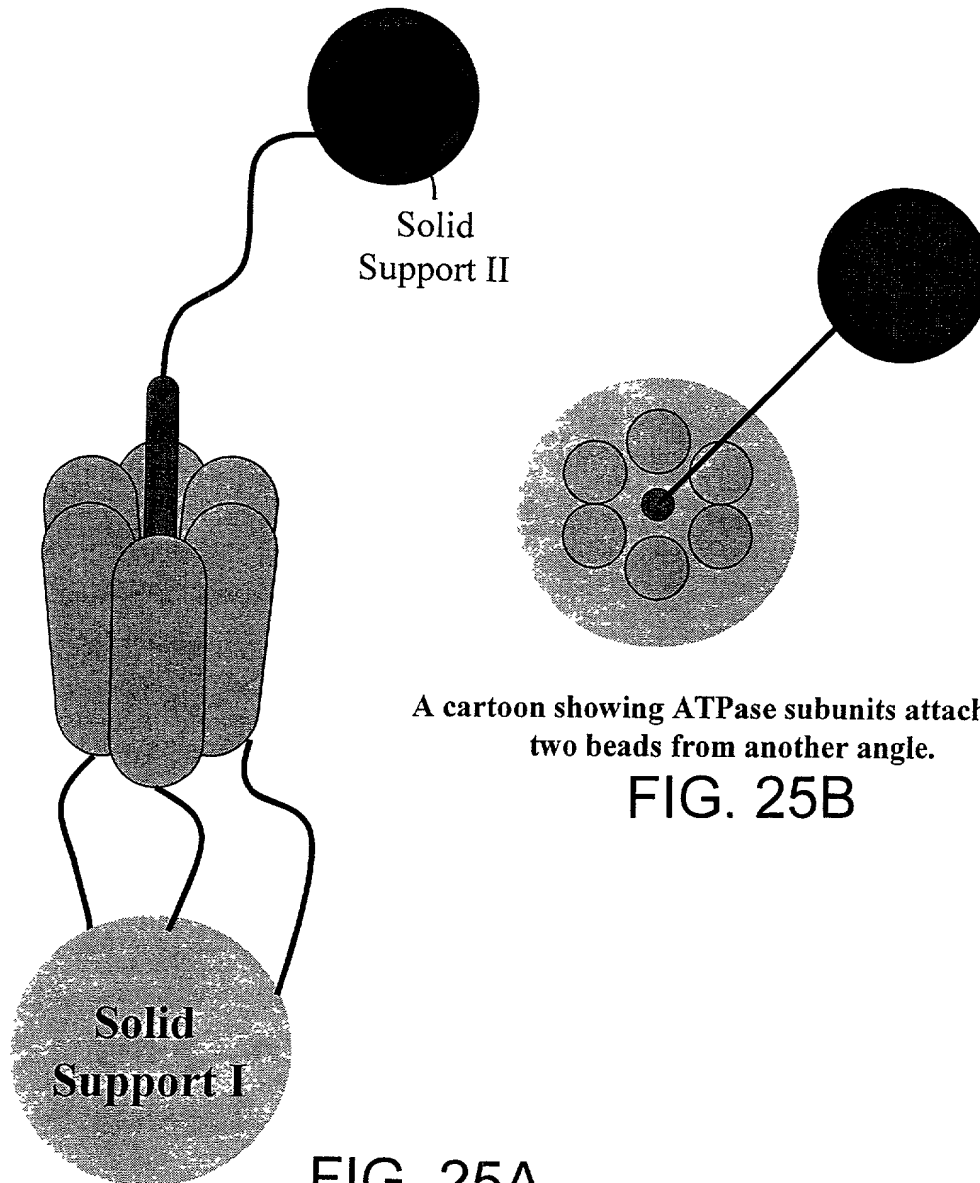
FIG. 24C



Flat platform base as Solid supports for attaching one of the subunits of ATPase

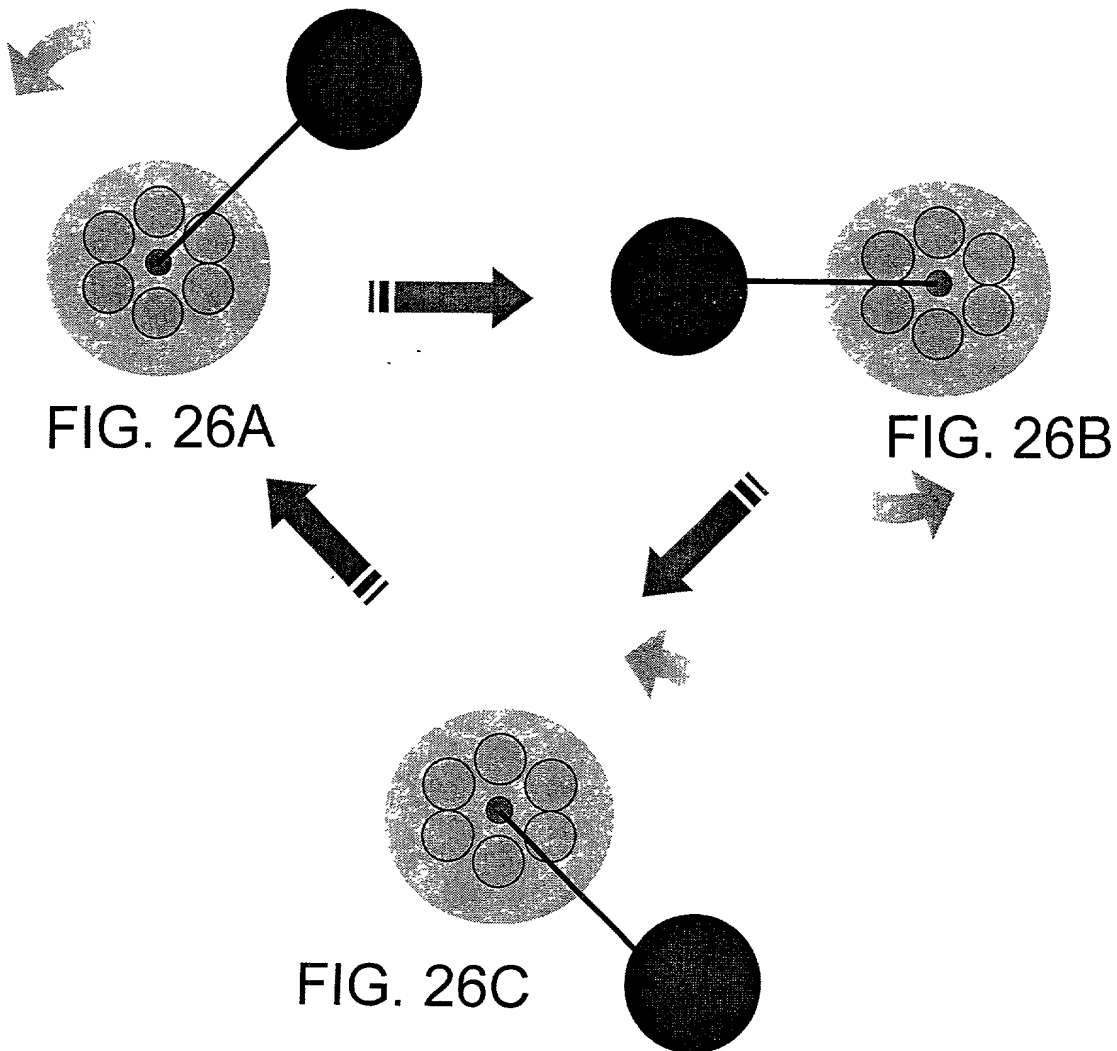
FIG. 24D

## ATPase based Fluid-Micromixers, Model I



One particular implementation of ATPase-based fluid-micromixers. The  $\gamma$ -subunit and the  $\alpha,\beta$ -subunit core are both attached to two different spherical beads.

A cartoon showing rotation of the two beads  
bound to ATPase (Model I micromixer)  
upon addition of ATP



## A cartoon showing multiple Model I Micromixers in action in a solution

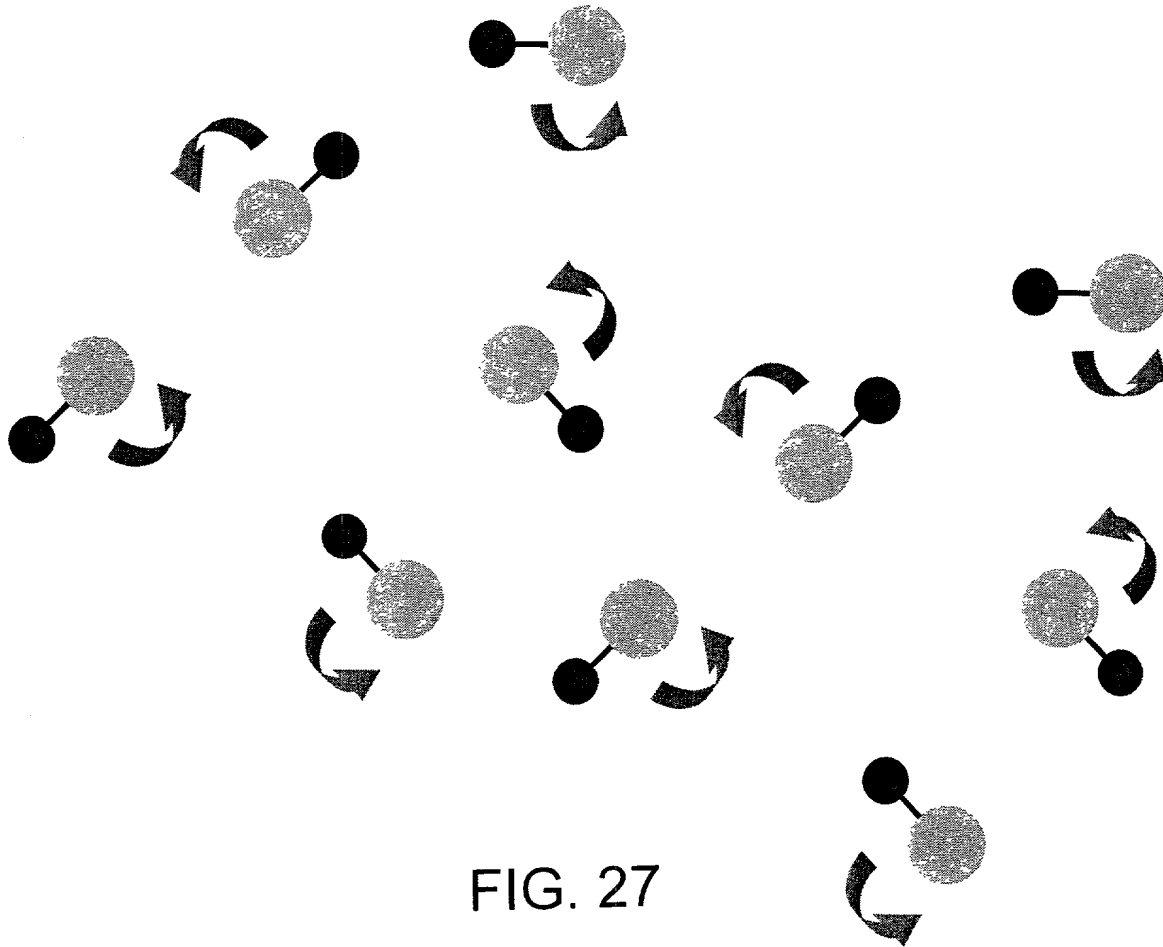
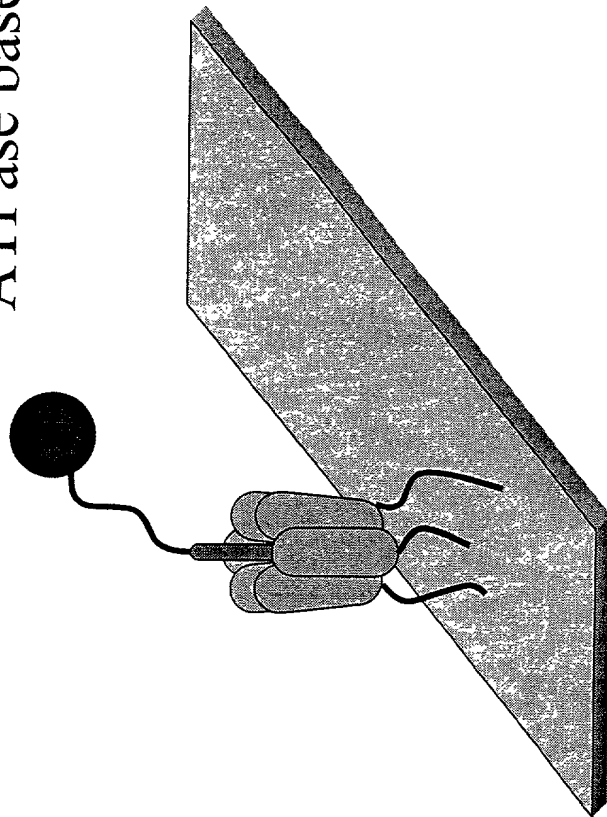


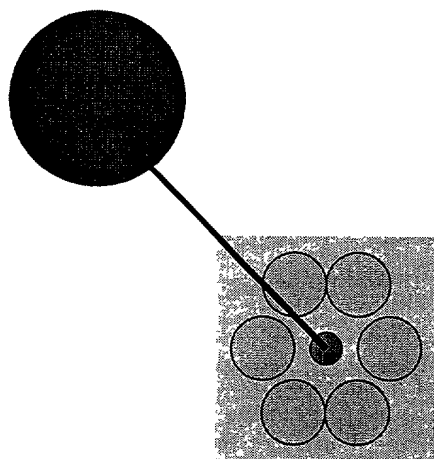
FIG. 27

# ATPase based Fluid-Micromixers, Model II



Another implementation of ATPase-based fluid-micromixers.  
The  $\gamma$ -subunit is attached to a spherical bead and the  $\alpha, \beta$ -subunit core is attached to a solid platform.

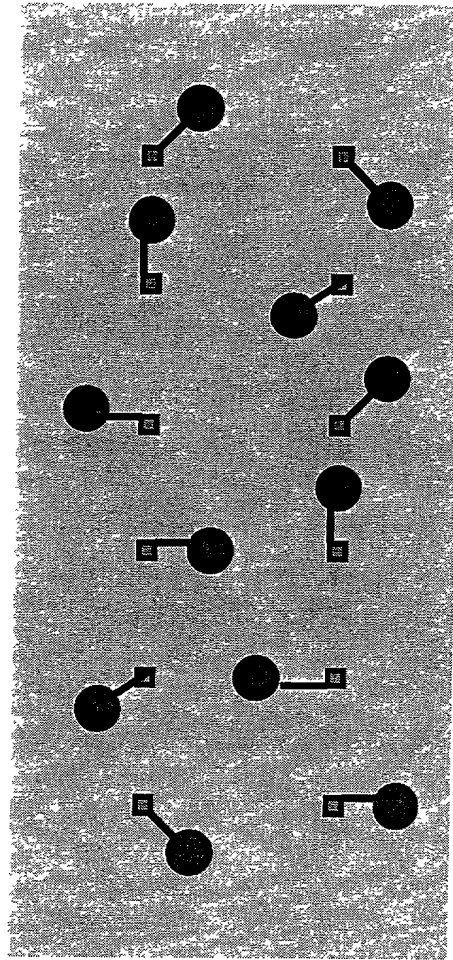
FIG. 28A



A cartoon showing ATPase subunits attached to two different surfaces from another angle.

FIG. 28B

## A cartoon showing multiple Model II Micromixers in action in a solution

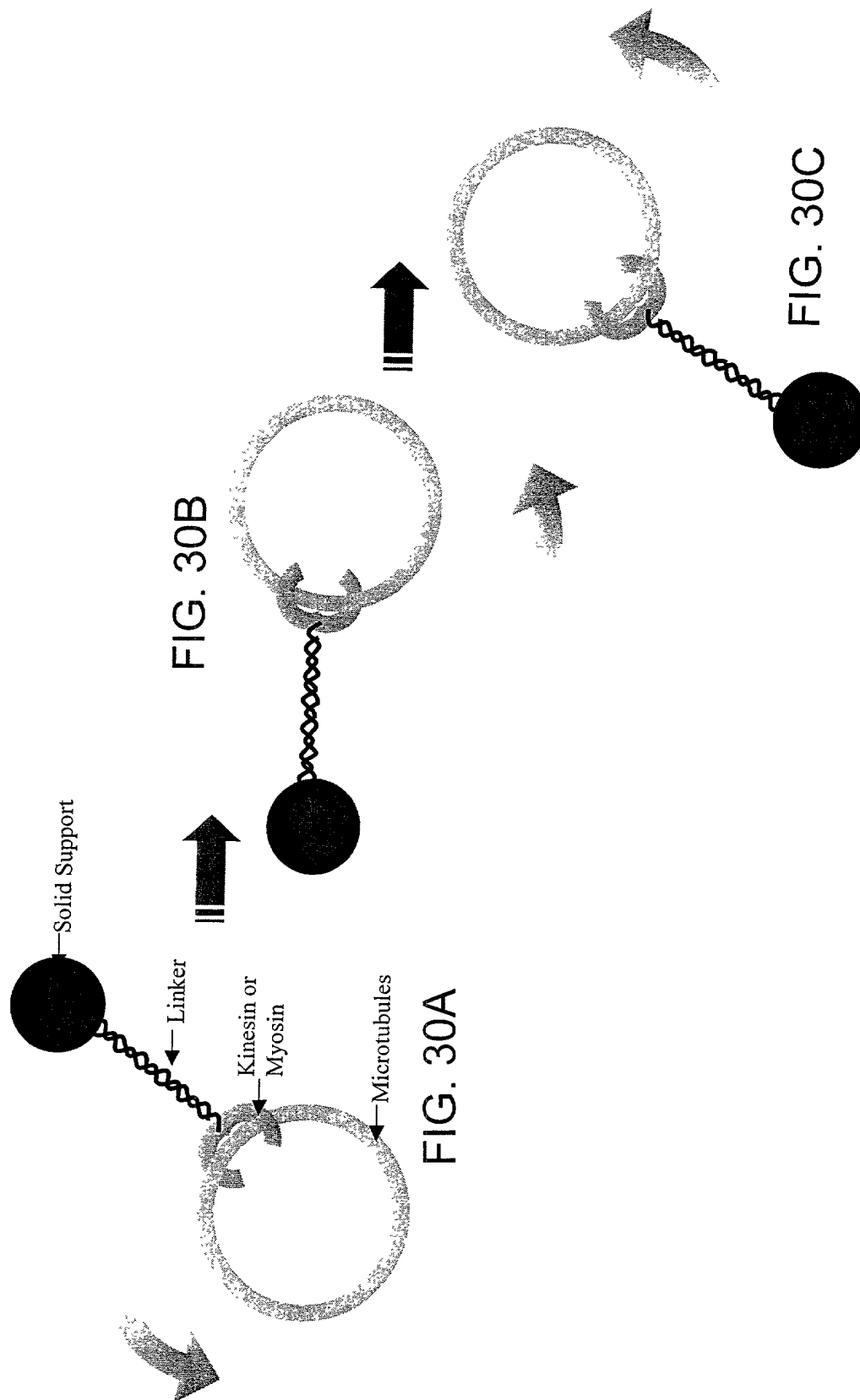


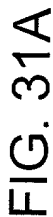
All the mixers are moving in a counter-clockwise direction

FIG. 29

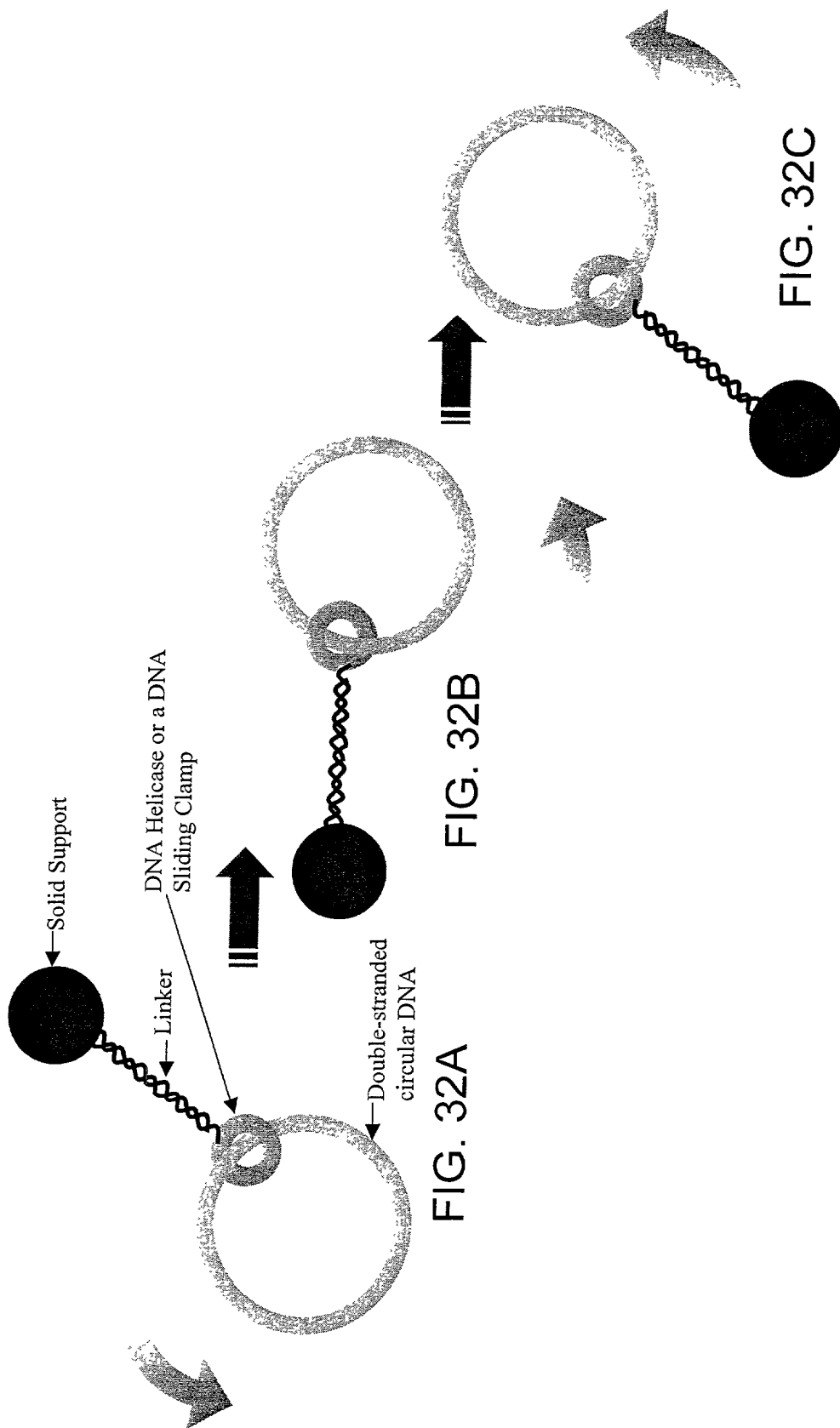


# Kinesin and Myosin based Fluid-Micromixers





# DNA Helicase and DNA Sliding Clamp based Fluid-Micromixers



# DNA Helicase (bound to Circular Triplex Forming Oligo (CTFO) or a Sliding Clamp) based Fluid-Micromixers

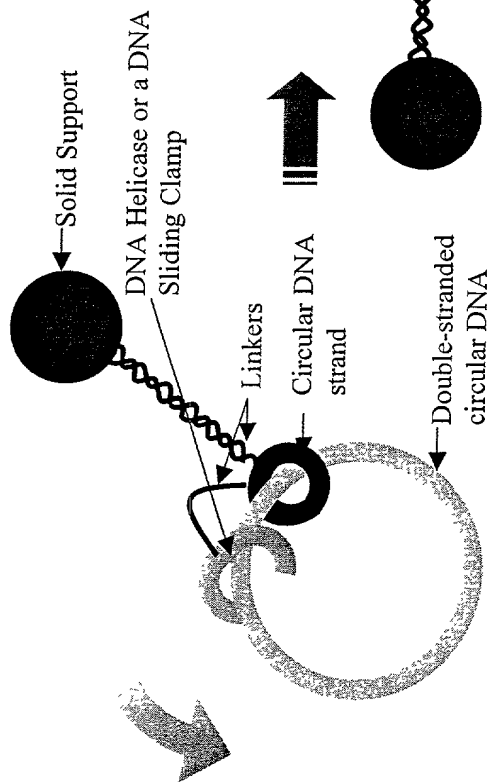


FIG. 33A

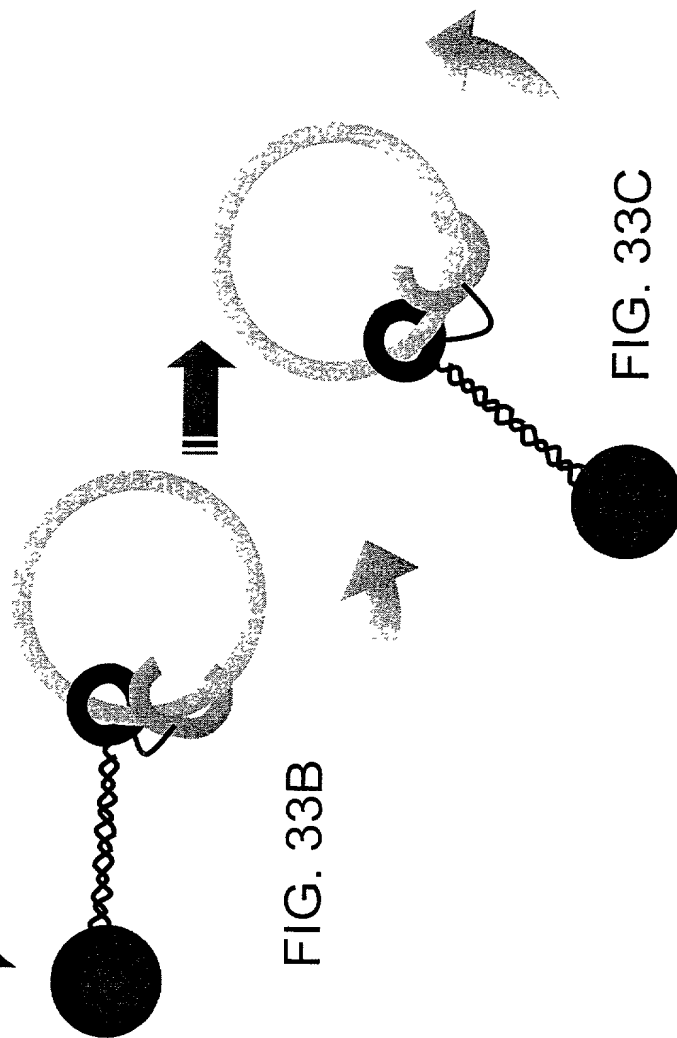
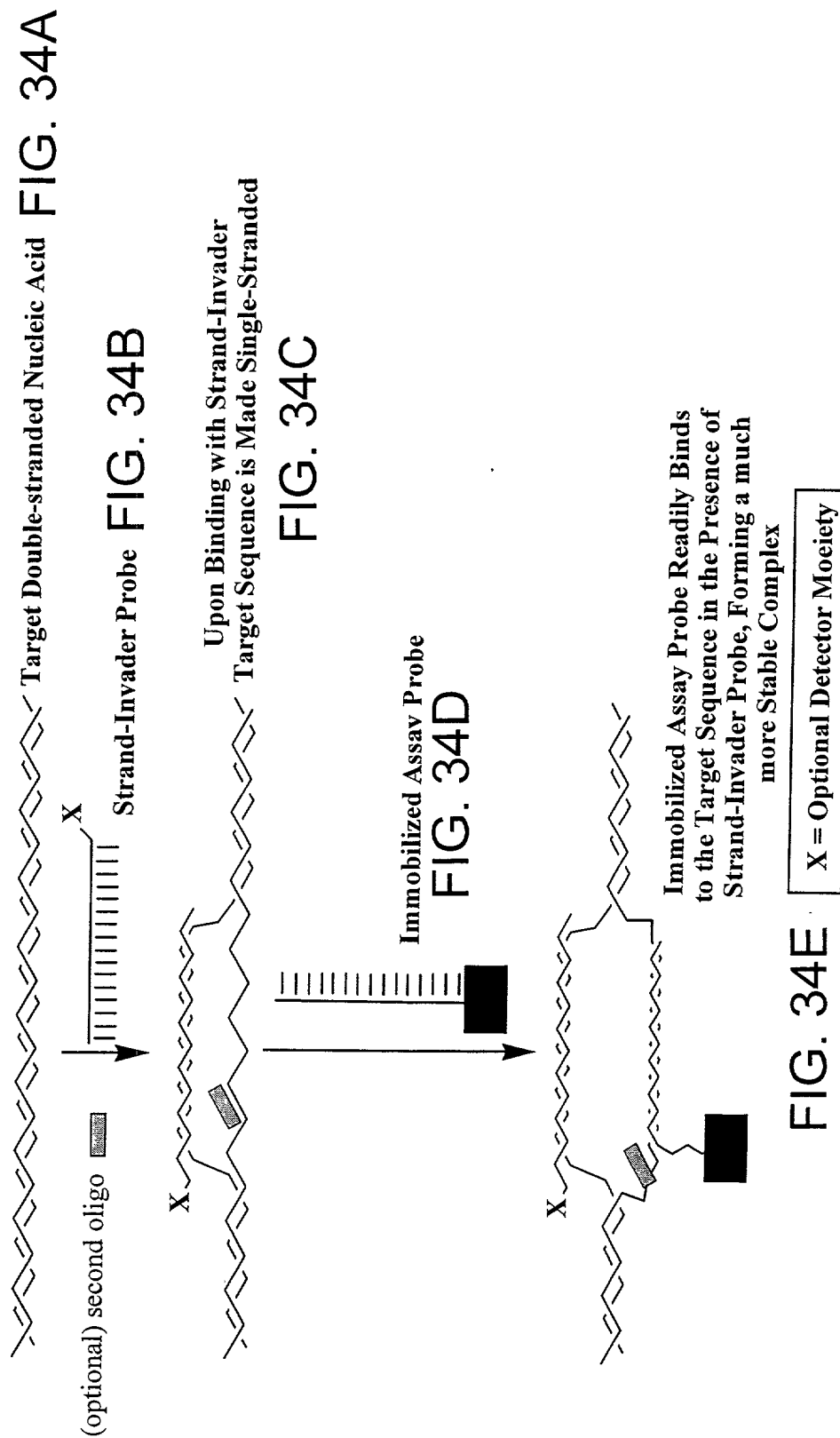


FIG. 33B

FIG. 33C

# Hybridization Enhancement Using Strand-Invader Molecules



A gasket/separator can be used in the current hybridization chambers to place two biochips facing each other in a single chamber for duplicate experiments.

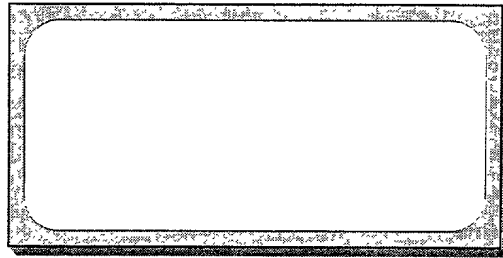


FIG. 35A

FIG. 35B

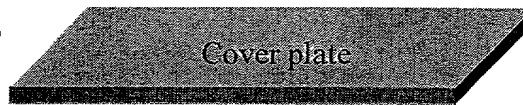


FIG. 35C



Biochip

FIG. 35D



Separator

FIG. 35E



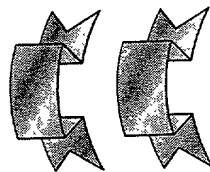
Biochip

FIG. 35F



Holder

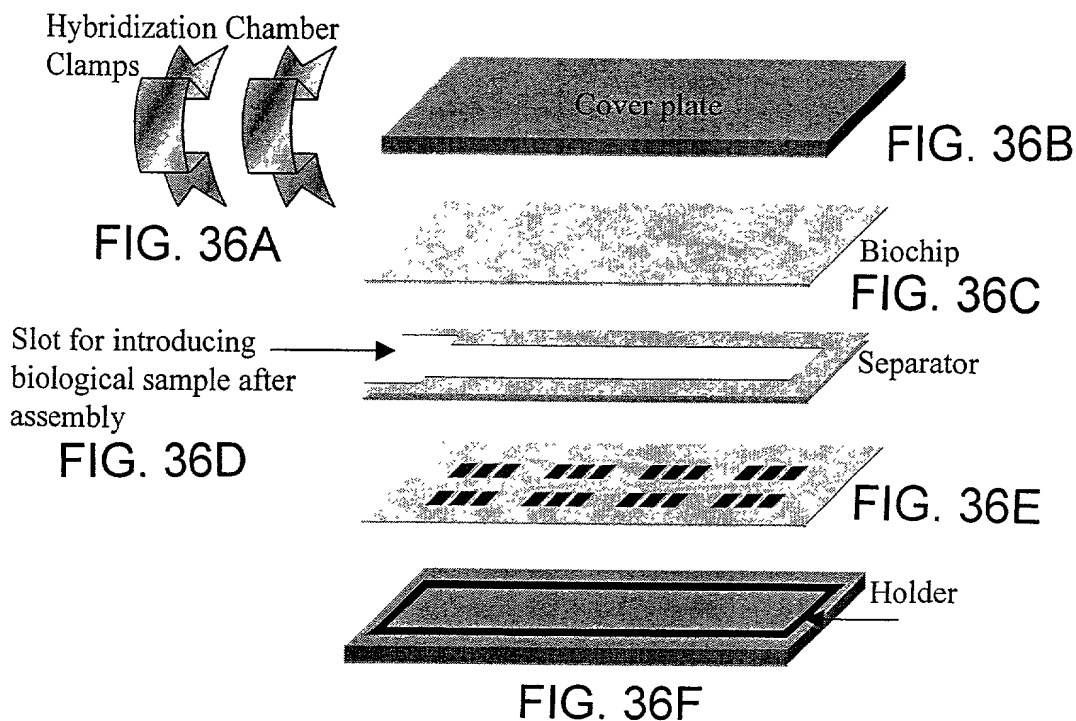
A sketch of one example of a new hybridization chamber.  
A hybridization chamber can be devised such that it fits  
two biochips.



Hybridization Chamber  
Clamps

FIG. 35G

- I A gasket/separator can be used in the current hybridization chambers to place two biochips facing each other in a single chamber for duplicate experiments.



- II The separator can also be built into the chamber.

